

# Selected Topics in Protein Structure and Function

Structural Basis of ABC Transporters

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**Faculté de médecine | Faculty of Medicine**

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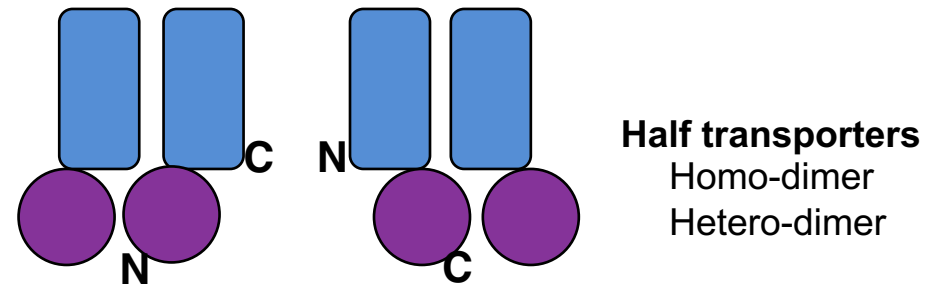
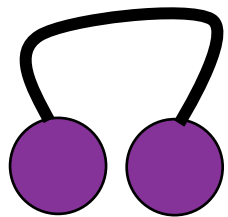
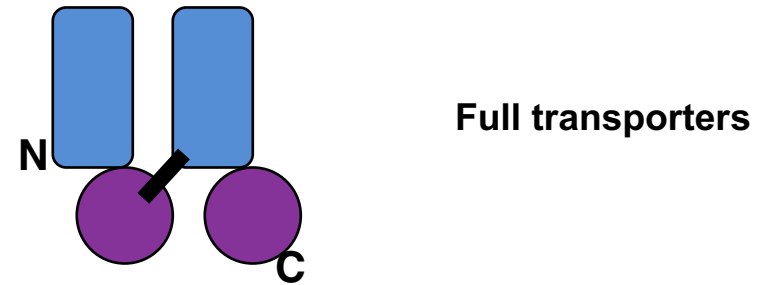
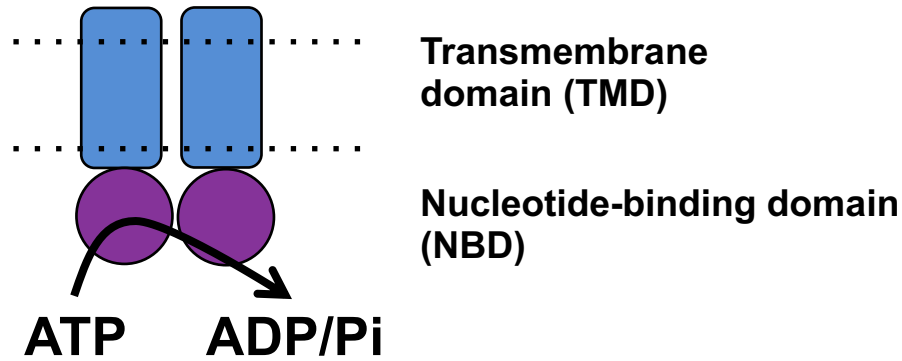
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# Structural basis: what do we do with membrane protein structures?

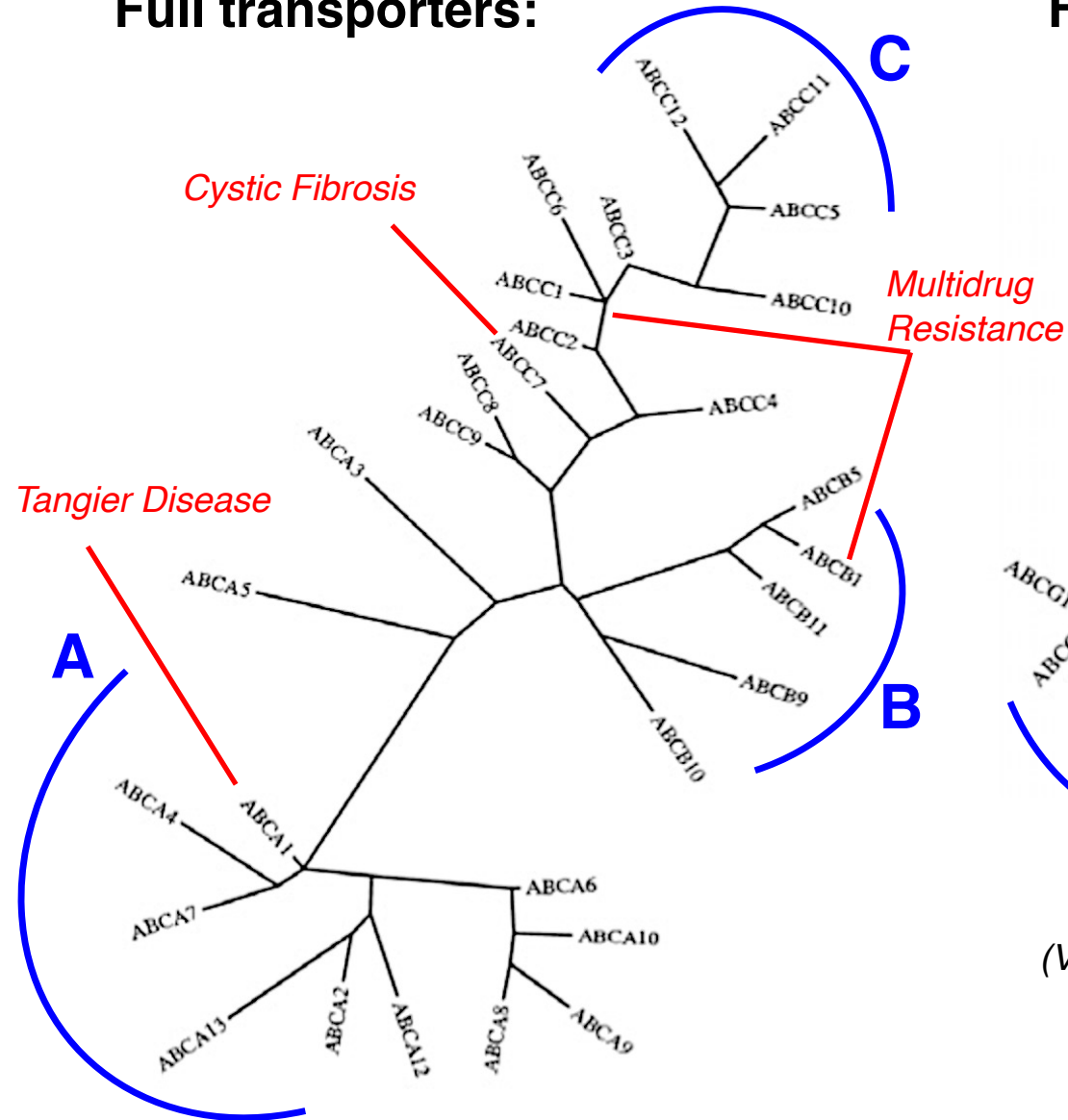
- 1. Protein-protein interactions**
- 2. Ligand recognition**
3. DNA and RNA binding
4. Hydrogen atoms critical for macromolecule functions
5. Kinetics
- 6. Enzymatic reaction**
7. Oncogenes and tumor suppressors
8. Drug Design
  
- 9. Lipid-protein interaction**
- 10. Structure-function crosstalk in the cellular membranes**
11. ...

# ATP-binding cassette (ABC) transporters (more than just ABC)

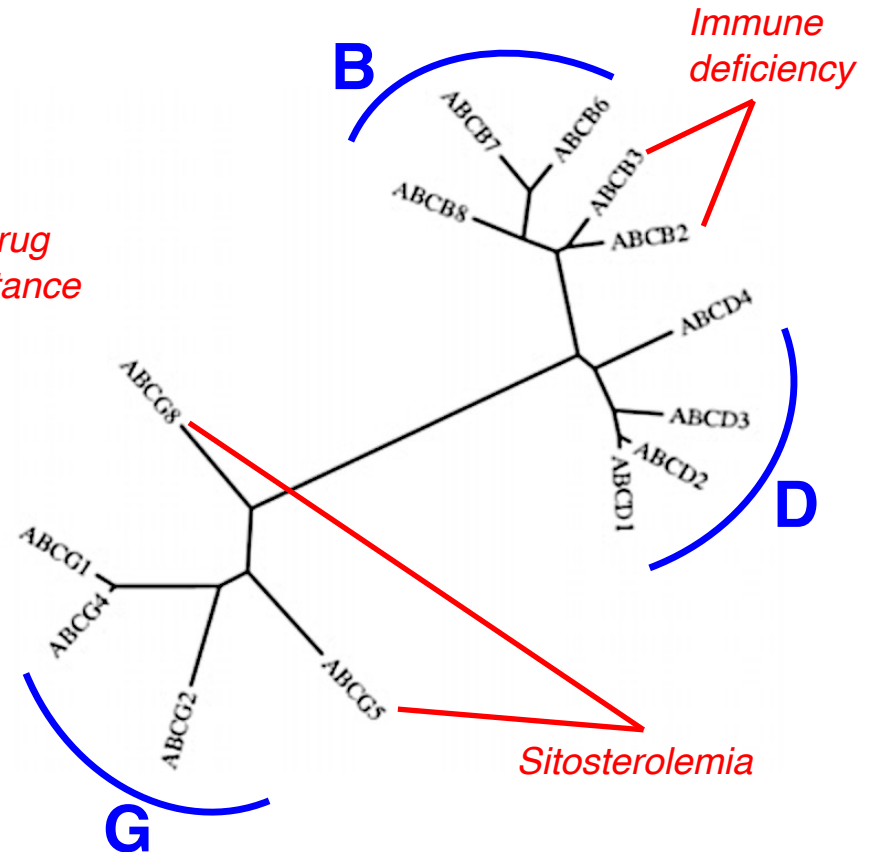


# ABC Transporters v.s. Human Diseases

## Full transporters:



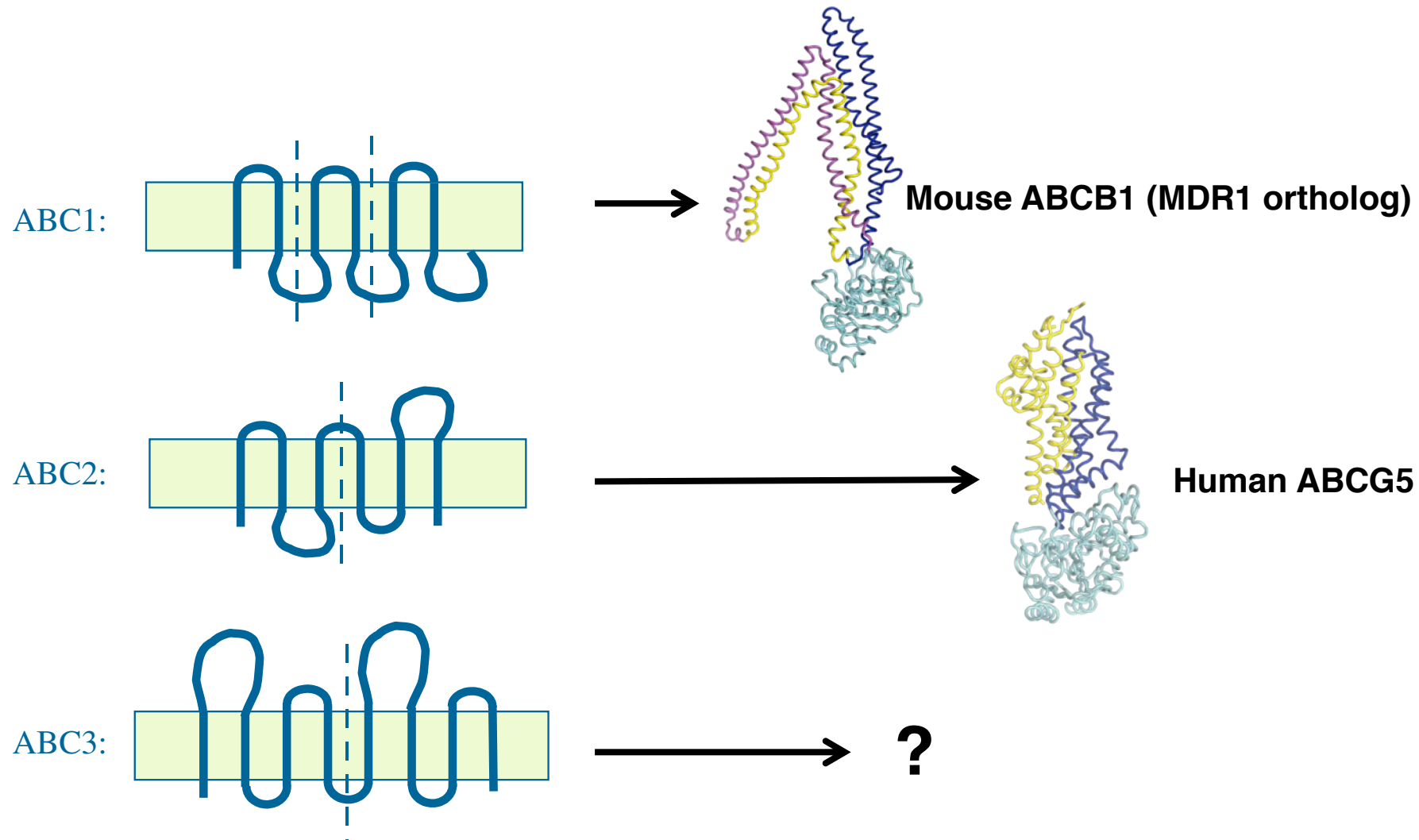
## Half transporters:



(Vishwakarma et al, Int J Sci Res, 2012)



# Evolutionary Origin of ABC Exporter TMD



(Wang et al, J Membr Biol, 2009)

# Human ABC proteins: 48 members, 44 transporters

## Transporters

- B, C, D
- A, G

## Non-transporters

- E, F

Table 1. Human ATP-binding cassette proteins.

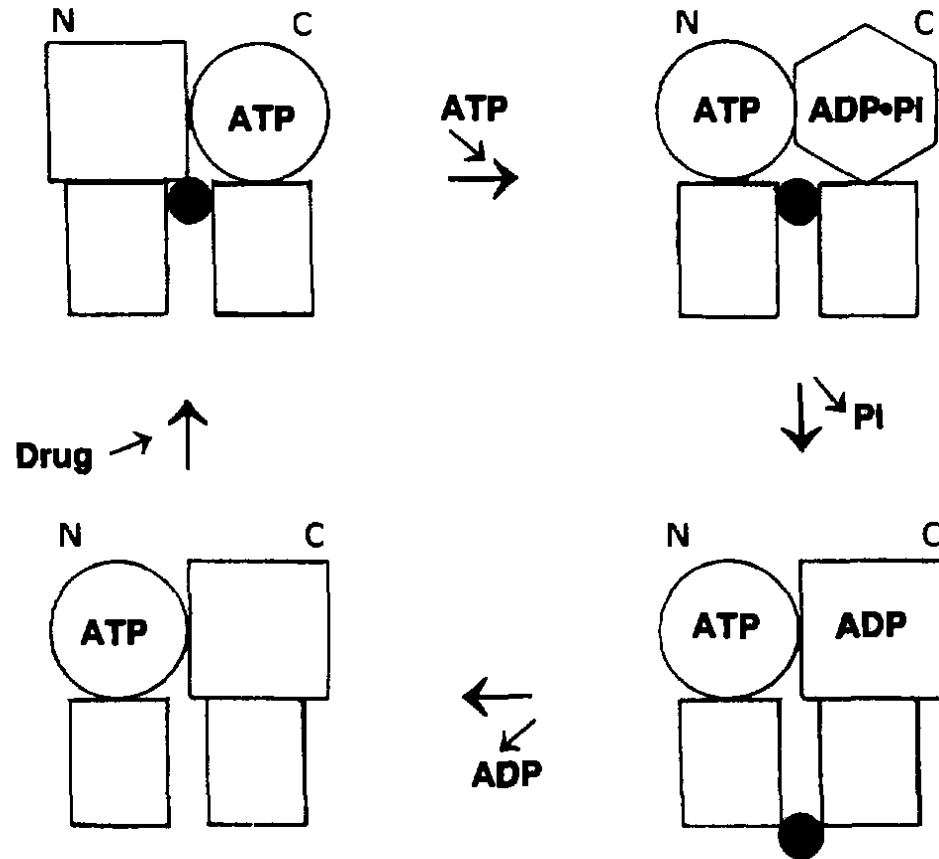
Subfamily	Human ABC proteins	Physiological role (known and probable)	Disease	Structure	Select citation(s)
ABC1	B: multidrug resistance, MDR, 11 members	ABC1	Efflux of xenobiotics	Kim and Chen 2018	Riordan et al. 1985
		ABC2	Peptide transport associated with antigen processing	Oldham et al. 2016	Deverson et al. 1990; Monaco et al. 1990; Spies et al. 1990; Trowsdale et al. 1990; Van der Blik et al. 1987
		ABC3			
		ABC4	Phospholipid excretion into bile		Progressive familial intrahepatic cholestasis type III
		ABC5	Efflux of xenobiotics		Allikmets et al. 1996
		ABC6	Porphyrin transport		Mitsuhashi et al. 2000
		ABC7	Transport substrate involved in the mitochondrial iron homeostasis	X-linked sideroblastic anemia with ataxia	Savary et al. 1997
		ABC8	Mitochondrial iron and glutathione export; efflux of xenobiotics		Allikmets et al. 1996
		ABC9	Peptide translocation to lysosomes		
		ABC10	Involved in heme biosynthesis		
		ABC11	Bile salt secretion into bile	Progressive familial intrahepatic cholestasis type II	Shintre et al. 2013; Zhang et al. 2000; Strautnieks et al. 1998
	C: multidrug resistance-associated protein, MRP, 12 members	ABCC1	Multispecific organic anion transport	Multidrug resistance	Martin et al. 2017a; Cole et al. 1992
		ABCC2	Renal and biliary elimination of organic anionic substrates	Dublin-Johnson syndrome	Büchler et al. 1996
		ABCC3	Organic anion transport		Kiuchi et al. 1998
		ABCC4	Nucleotide transport; antiviral drug efflux		Kool et al. 1997
		ABCC5	Nucleotide and glutamate conjugate transport		Jedlitschky et al. 2000; Wijnholds et al. 2000
		ABCC6	Transport of organic anions	Pseudoxanthoma elasticum	Kuss et al. 1998
		ABCC7	Epithelial chloride channel	Cystic fibrosis; congenital bilateral absence of the vas deferens	Liu et al. 2017; Riordan et al. 1989
	D: adrenoleukodystrophy-related protein, ALD, 4 members	ABCC8	Modulation of associated potassium channels	Hyperinsulinemic hypoglycemia of infancy	Martin et al. 2017a; Aguilar-Bryan et al. 1995
		ABCC9		Cantu syndrome	Chutkow et al. 1996
		ABCC10	Efflux of xenobiotics	Resistance to anticancer and antiviral nucleoside based drugs	Allikmets et al. 1996
		ABCC11	Anionic hydrophobic solute transport		Lagasse and Clerc 1988
		ABCC12	Unknown		Tammur et al. 2001
		ABCD1	Long and very long chain fatty acid transport	Adrenoleukodystrophy	Mosser et al. 1993
		ABCD2			Holzinger et al. 1999
ABC2	ABC2 A: 12 members	ABCD3	Branched chain fatty acid transport		Zellweger syndrome
		ABCD4	Possible role in vitamin B12 transport		Kamijo et al. 1990
		ABCA1	Cholesterol and phospholipid transport	Tangier disease; familial high-density lipoprotein deficiency	Holzinger et al. 1997; Luciani et al. 1994
		ABCA2	Phospholipid transport		
		ABCA3	Phospholipid transport	Neonatal surfactant deficiency	Connors et al. 1997
		ABCA4	Transport of retinoid	Stargardt macular degeneration; cone-rod dystrophy	Allikmets et al. 1997
		ABCA5	Nucleotide and glutamate conjugate transport		Arnould et al. 2002
		ABCA6	Role in macrophage lipid homeostasis		Kaminski et al. 2001
		ABCA7	Phospholipid and sphingolipid transport		Kaminski et al. 2000
		ABCA8	Cholesterol and taurocholate transport		Arnould et al. 2002
		ABCA9	Role in macrophage lipid homeostasis		Piehl et al. 2002
	G: five members	ABCA10	Role in macrophage lipid homeostasis		Wenzel et al. 2003
		ABCA12	Sphingolipid transport	Harlequin ichthyosis	Annino et al. 2002
		ABCA13	Unknown		Prades et al. 2002
		ABCG1	Cholesterol and phospholipid transport		Chen et al. 1996; Savary et al. 1996
		ABCG2	Efflux of xenobiotics	Multidrug resistance	Taylor et al. 2017; Allikmets et al. 1998; Doyle et al. 1998; Miyake et al. 1999
		ABCG4	Cholesterol transport		Annino et al. 2001; Oldfield et al. 2002
		ABCG5	Cholesterol and plant sterol efflux	β-Sitosterolemia	Berge et al. 2000
	E: one member	ABCG8			
		ABCE1	Role in translation initiation and ribosome recycling		Preis et al. 2014; Shao et al. 2016 <sup>1,2</sup> ; Wolkoff et al. 1985
ABC3	F: three members	ABCF1	Regulation of innate immune response		Richard et al. 1998
		ABCF2	Role in cell volume regulation		Allikmets et al. 1996
		ABCF3	Probable role in cell proliferation		

Note: The 48 human ABC proteins from the subfamilies ABCA-G can be classified into two groups. Physiological function and disease phenotypes were obtained from [www.genecards.org](http://www.genecards.org) and <http://yeast>, and rabbit homologs, respectively.

(Xavier et al, BCB, 2019)

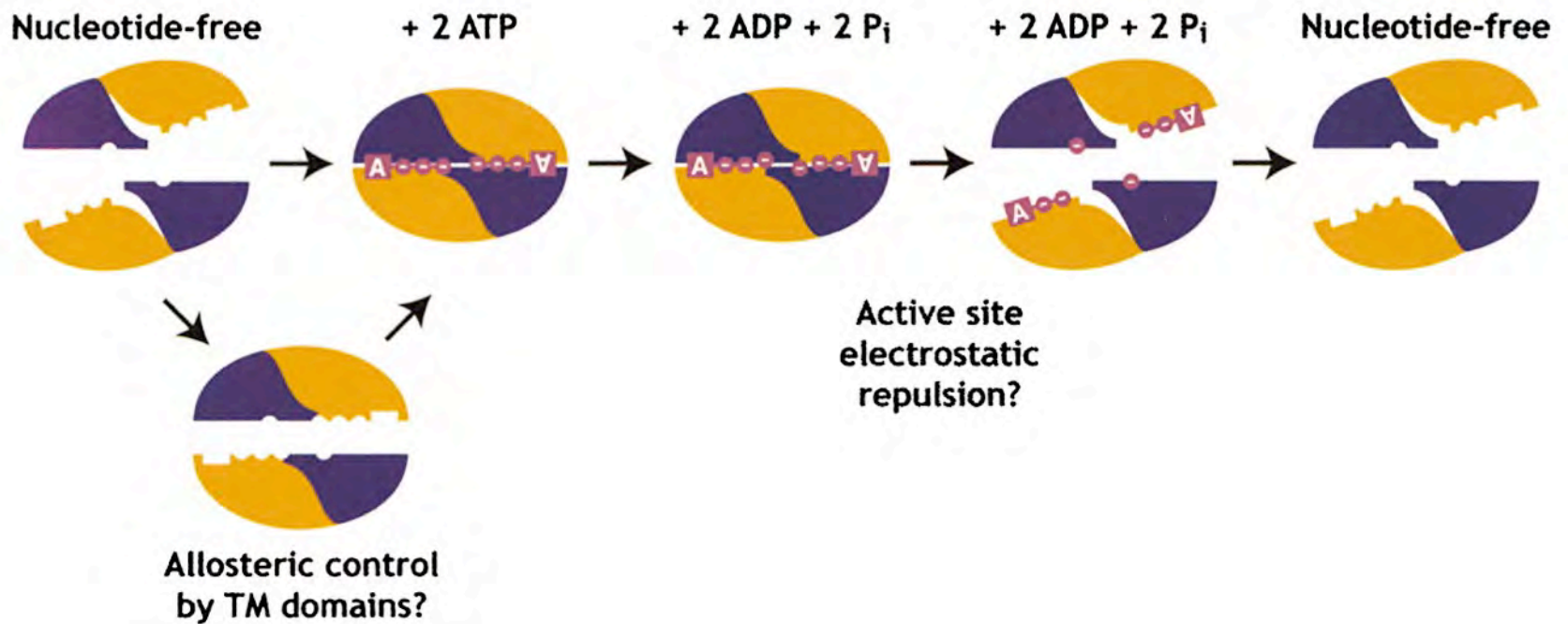
909) and Zheng et al. (2013). id 1, which indicate bovine,

# ABC: alternating catalytic cycle



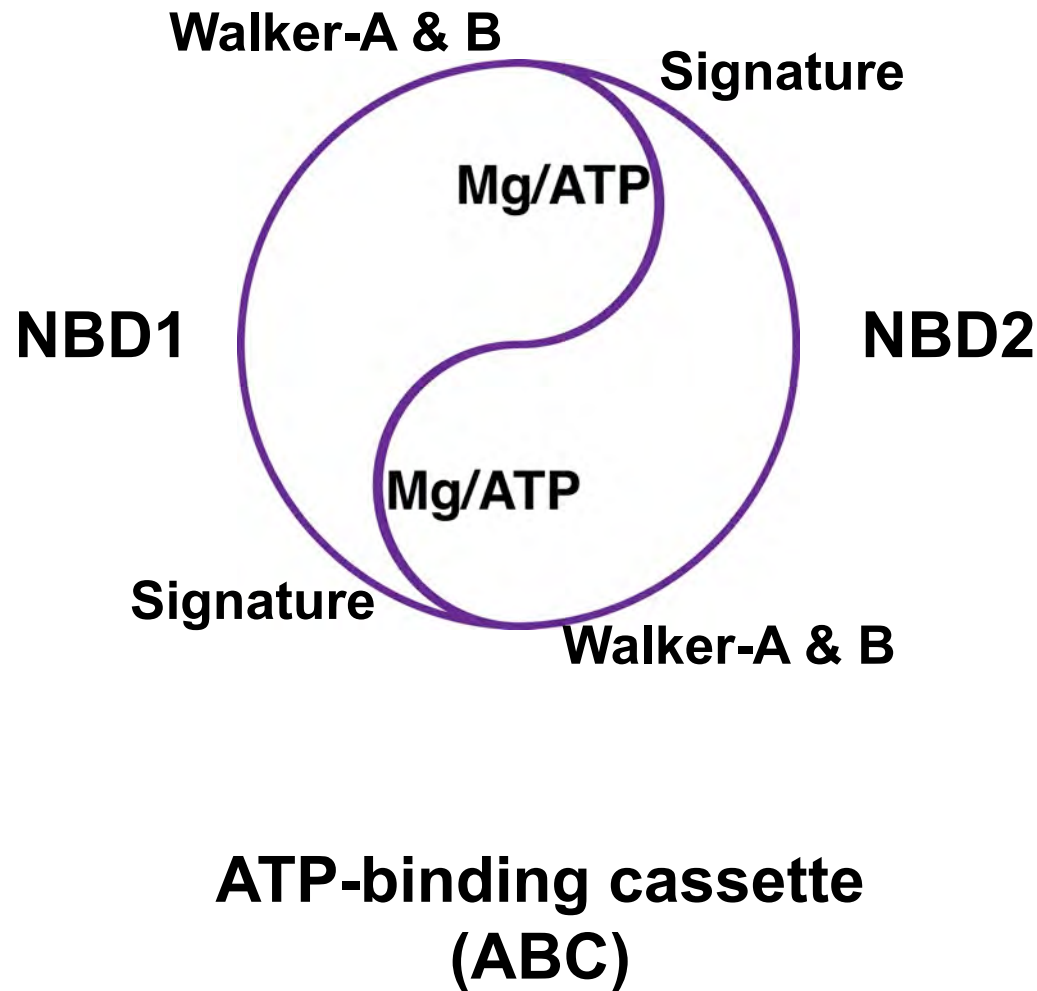
(Senior et al, FEBS Lett, 1995)

# ABC: ATP sandwich model

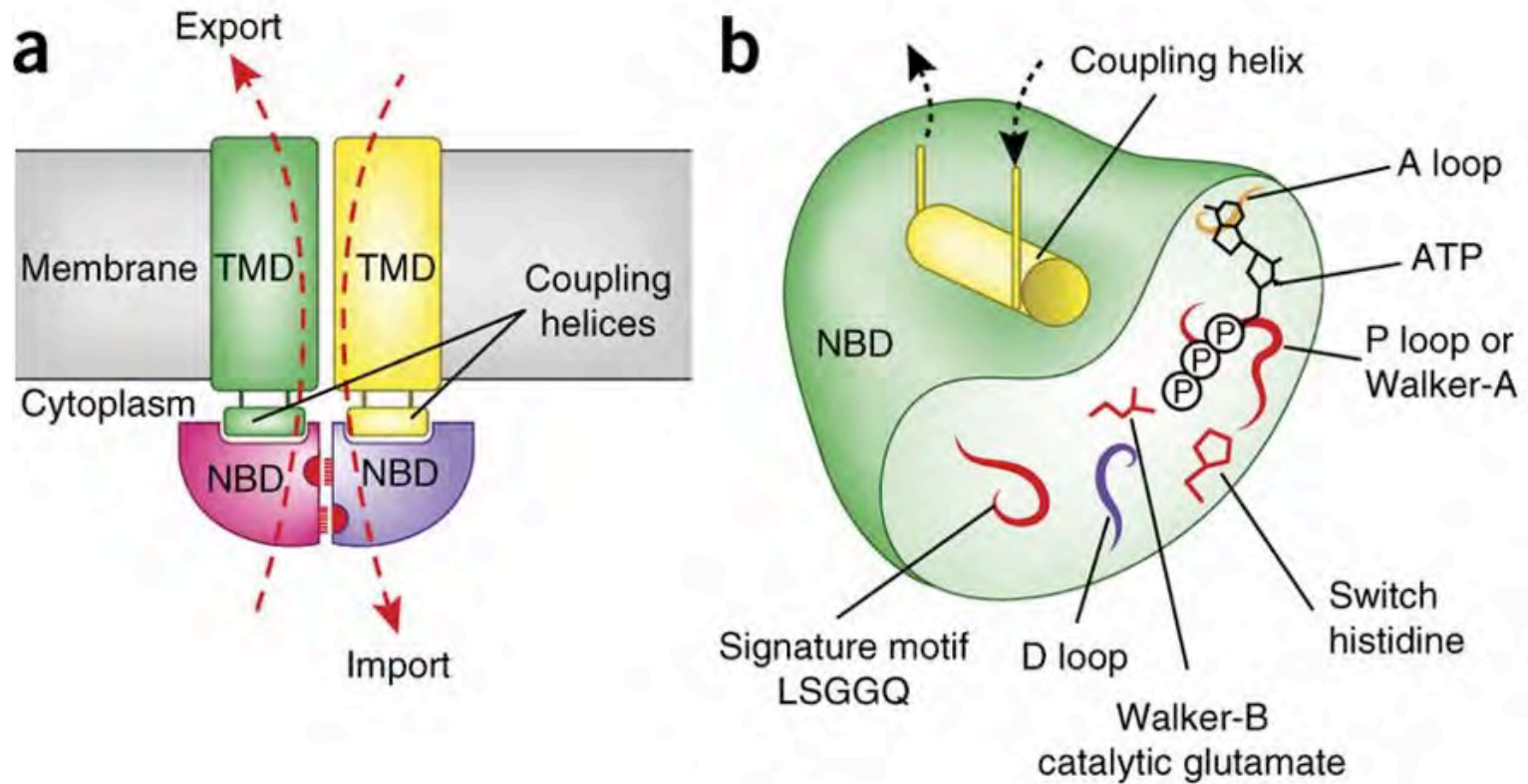


(Smith et al, Mol Cell, 2002)

# ABC coupled transport: a simple idea

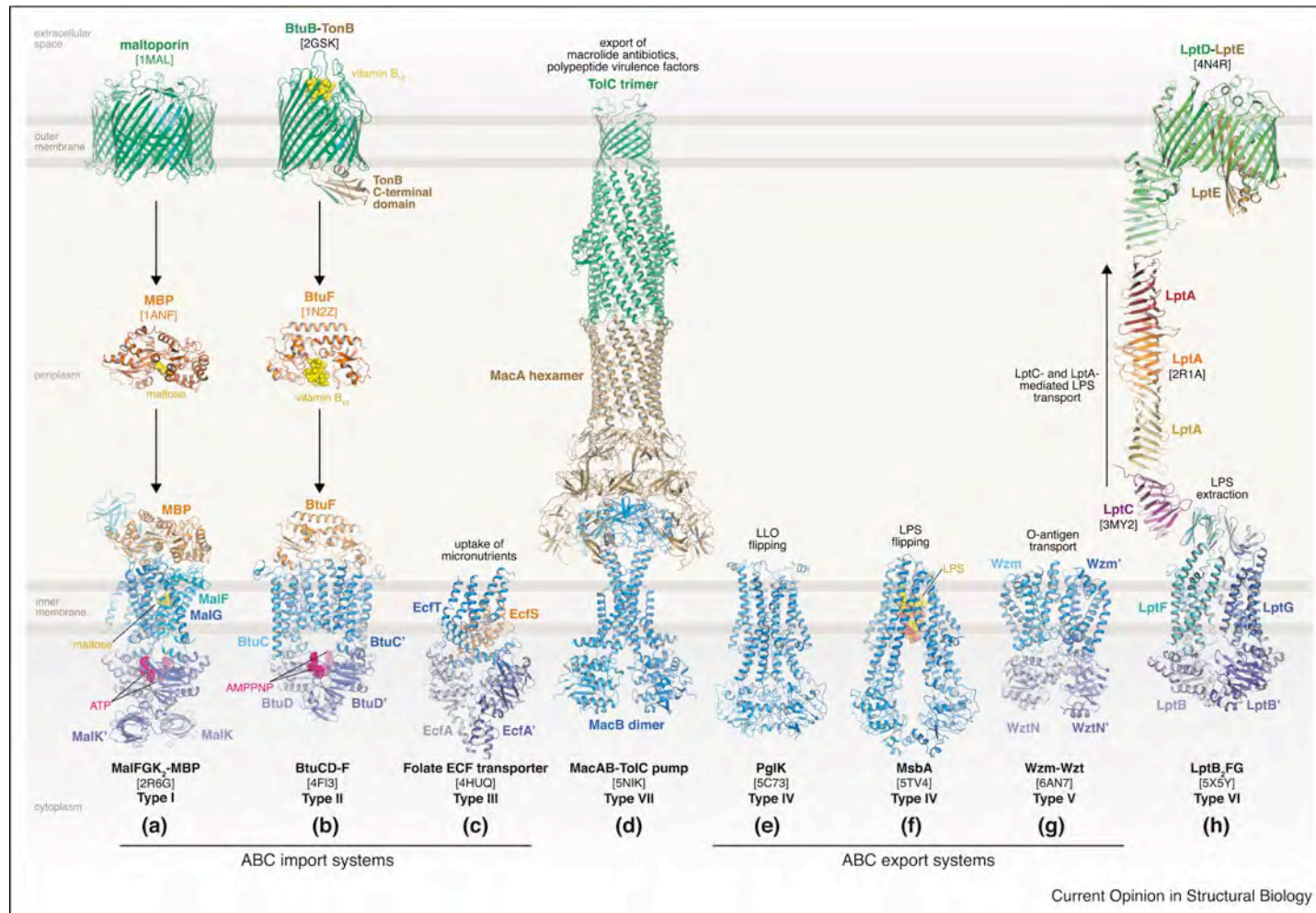


# ABC coupled transport: a simple idea



*(Locher, Nat Struct Mol Biol, 2016)*

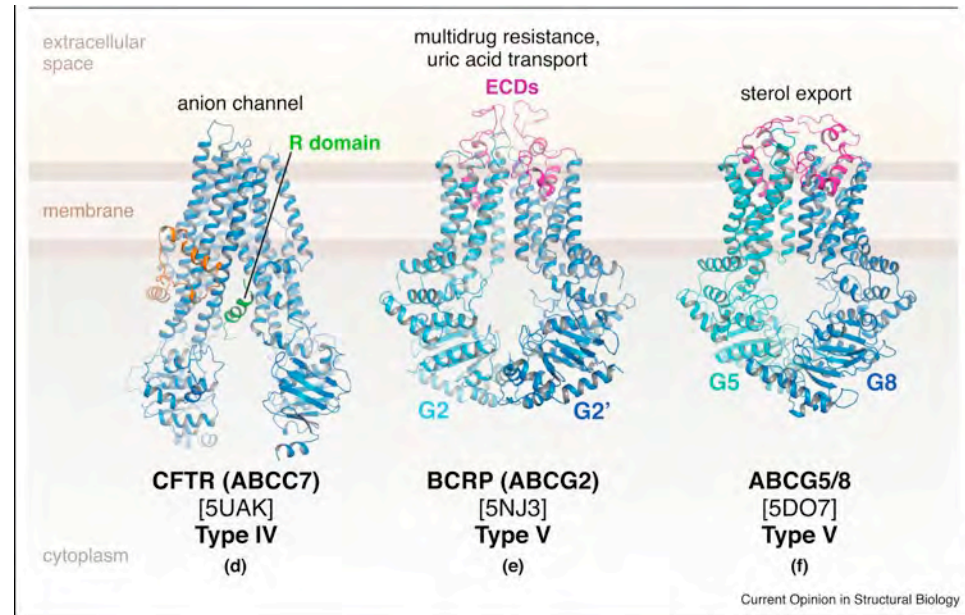
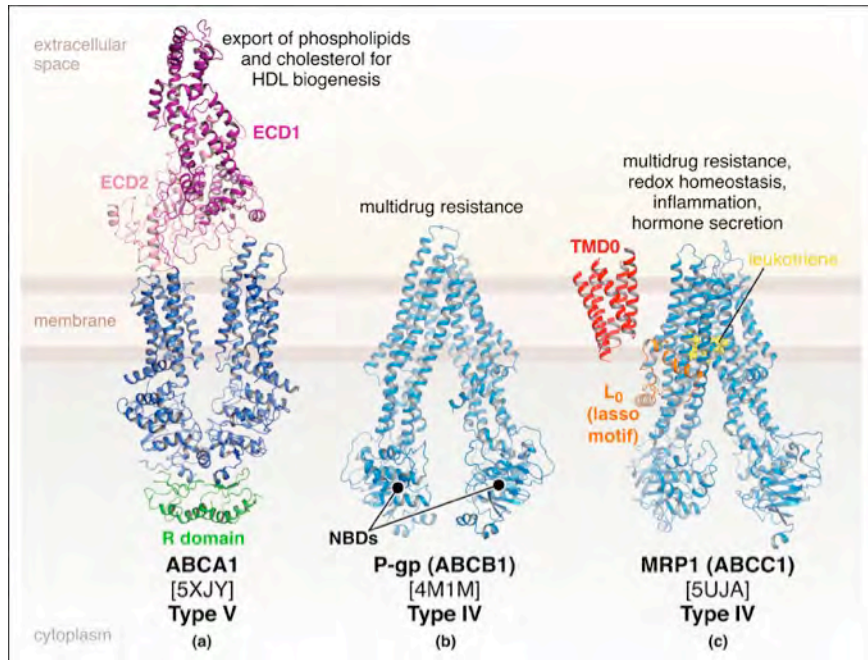
# Structural diversity: bacterial point of view



(Thomas & Tampé, Curr Opin Struct Biol, 2018)



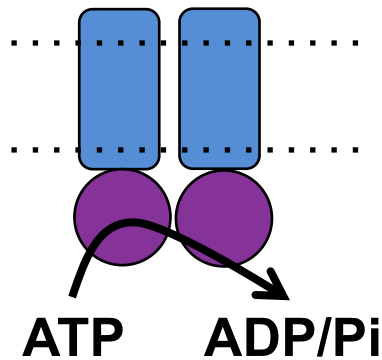
# Structural diversity: mammalian point of view



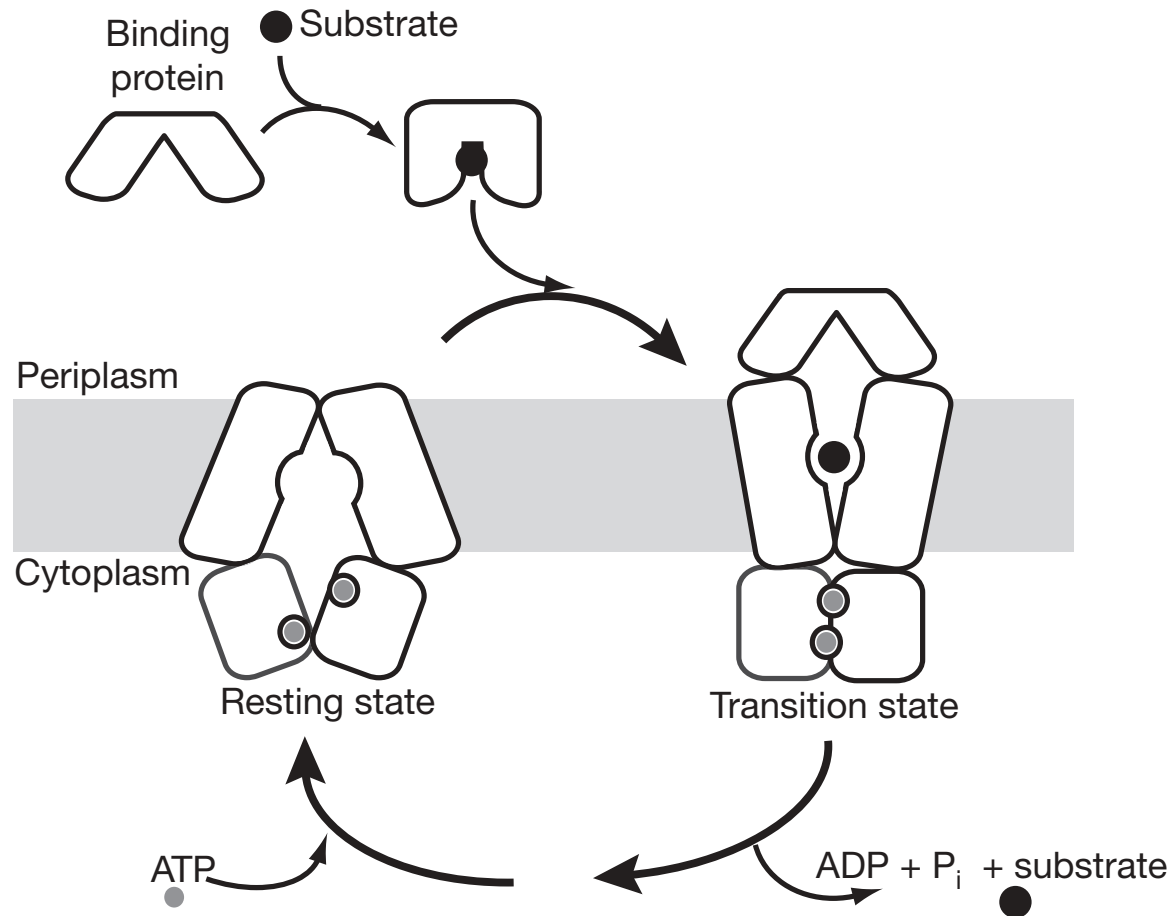
(Thomas & Tampé, Curr Opin Struct Biol, 2018)



## ABC and ATP usage are part of story!

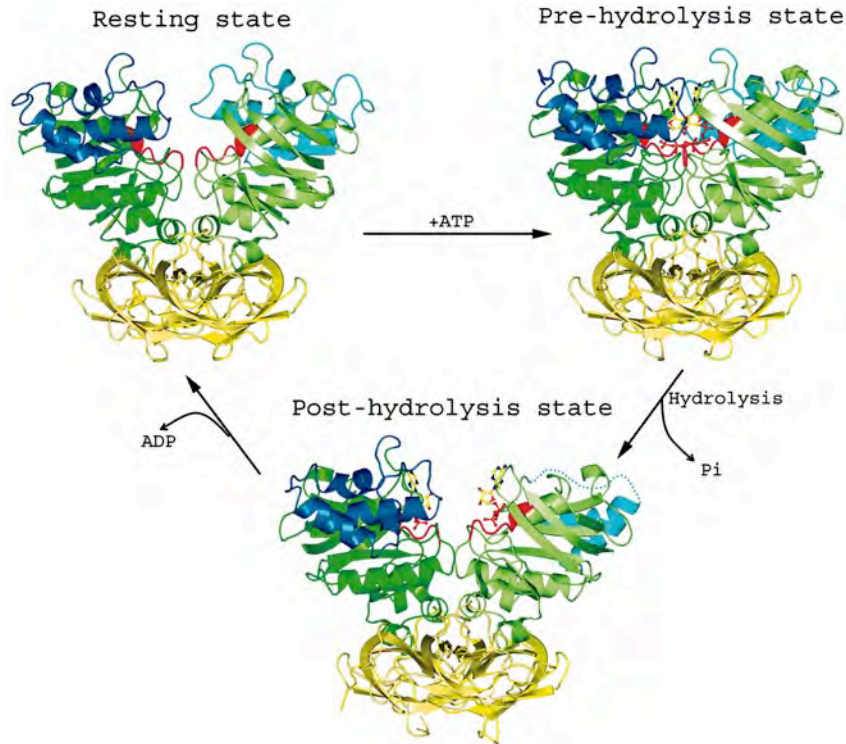


# Maltose transporter: a bacterial importer

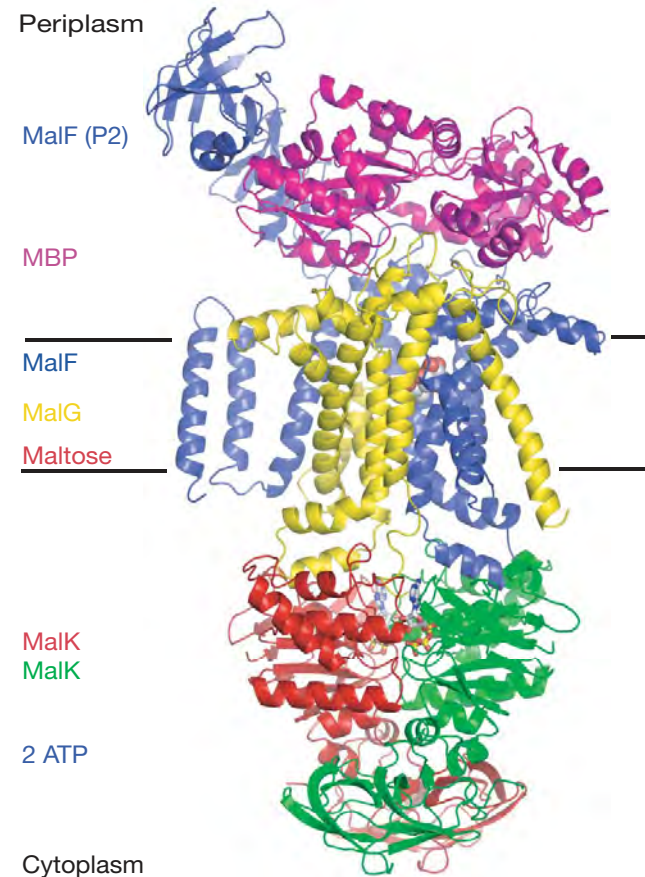


*(Olham et al, Nature, 2007)*

# Maltose transporter: a bacterial importer

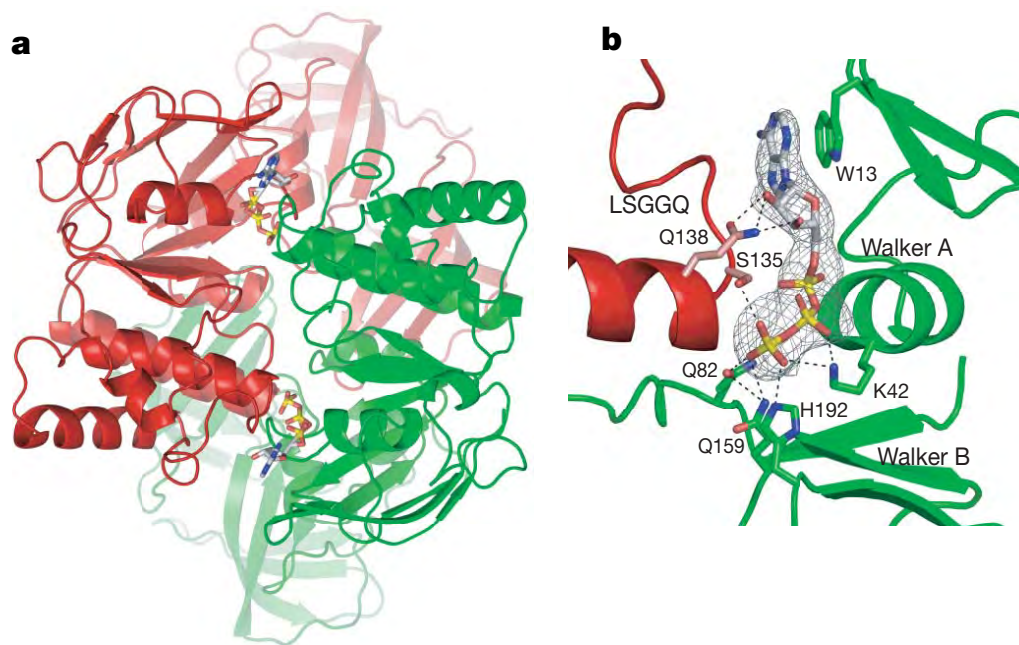


(Yu et al, PNAS, 2005)



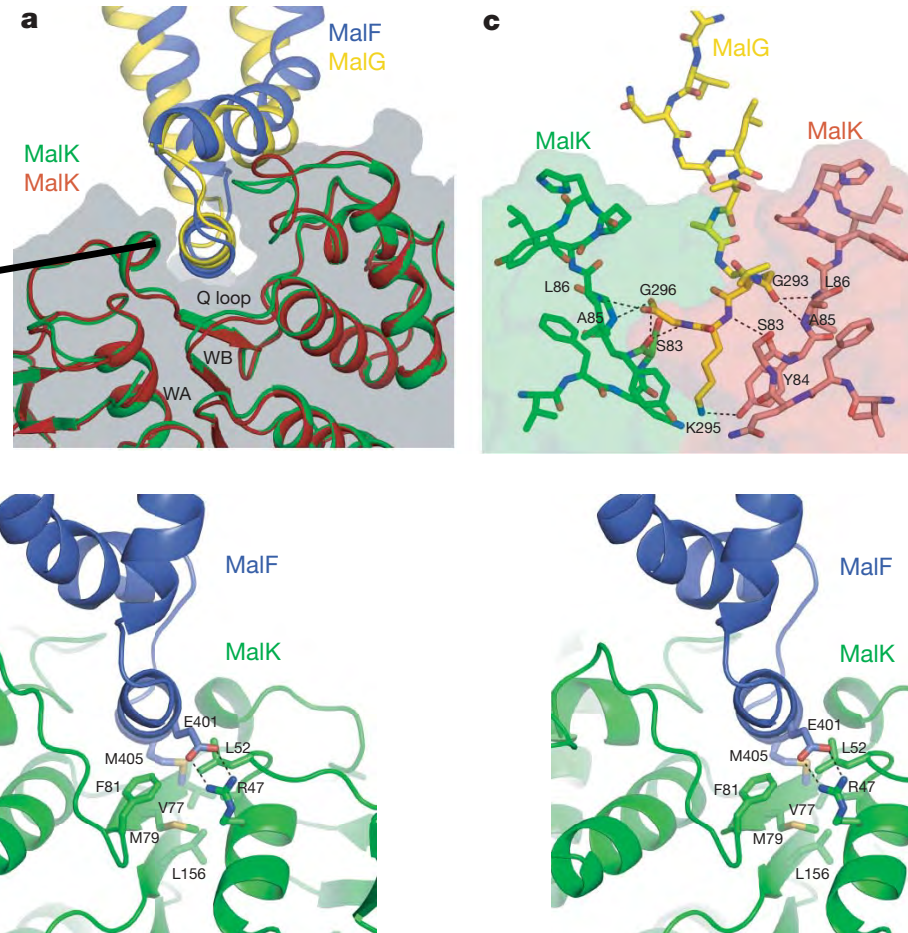
(Olham et al, Nature, 2007)

# Maltose transporter: ATP binding



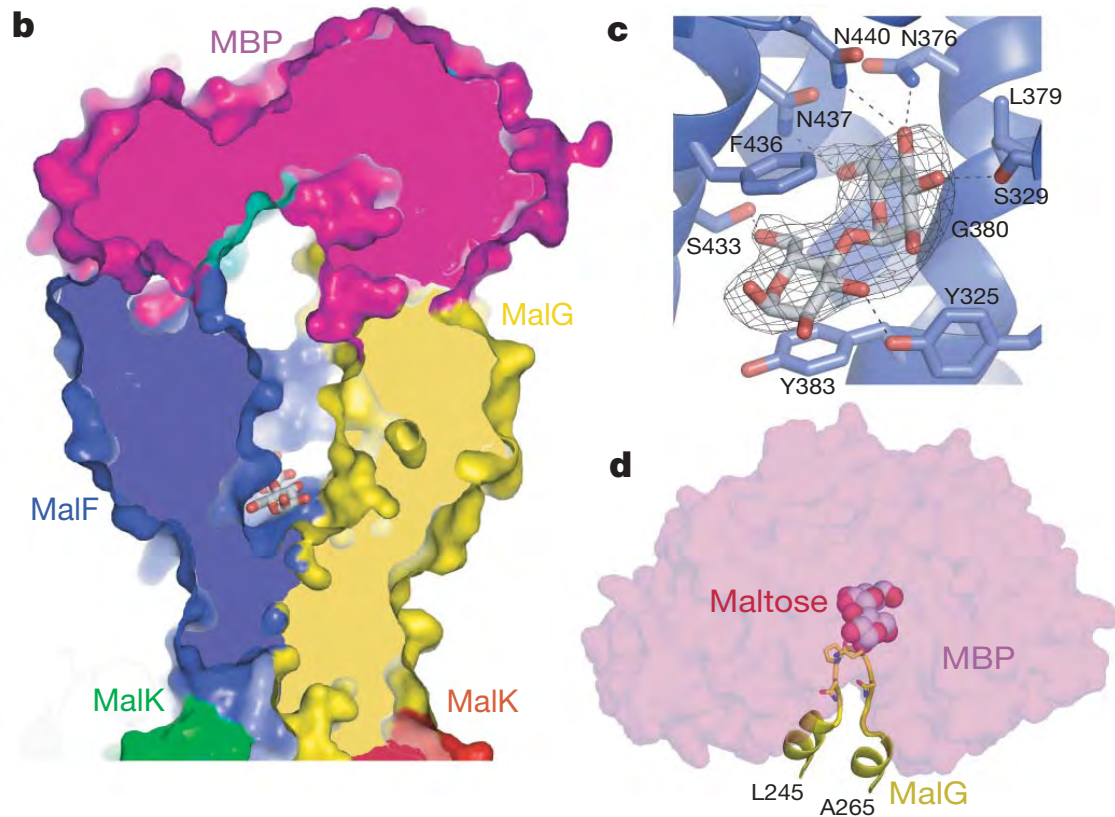
**Figure 2 | Ribbon diagram of the MalK subunits with bound ATP.** **a**, The MalK dimer viewed down the local twofold axis. The two subunits are coloured in red and green. The ATP is represented in ball-and-stick model. **b**, The ATP-binding site, showing that residues from both MalK subunits are making contact with the ATP. Interacting residues and associated hydrogen bonds (black dashed lines) from residues in the Walker A, Walker B and LSGGQ motifs, and from H192 and Q159, to ATP are indicated. A positive  $F_o - F_c$  electron density (contoured at  $1.5\sigma$ ) obtained with ATP omitted in the structure factor calculation is also shown.

# Maltose transporter: first molecular view of TMD-NBD interaction

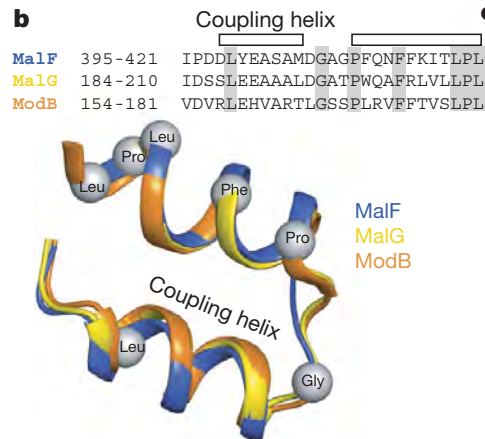
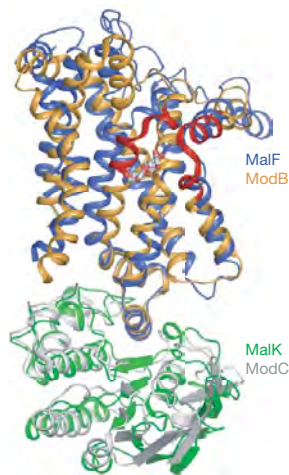
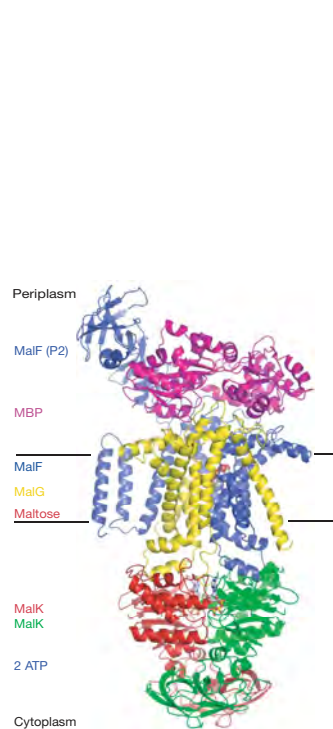




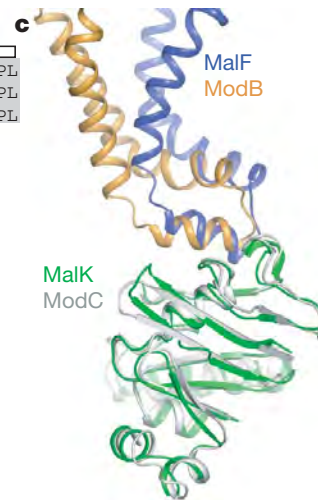
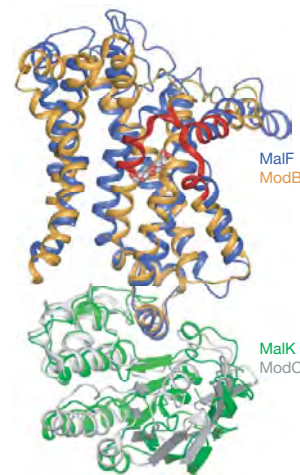
# Maltose transporter: transport-substrate binding



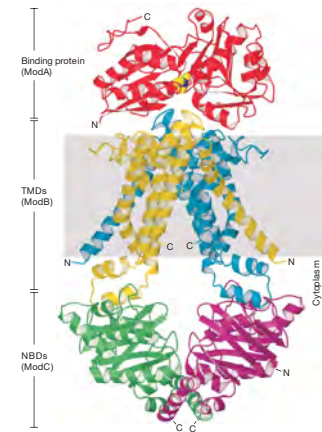
# Maltose transporter v.s. other importer(s)



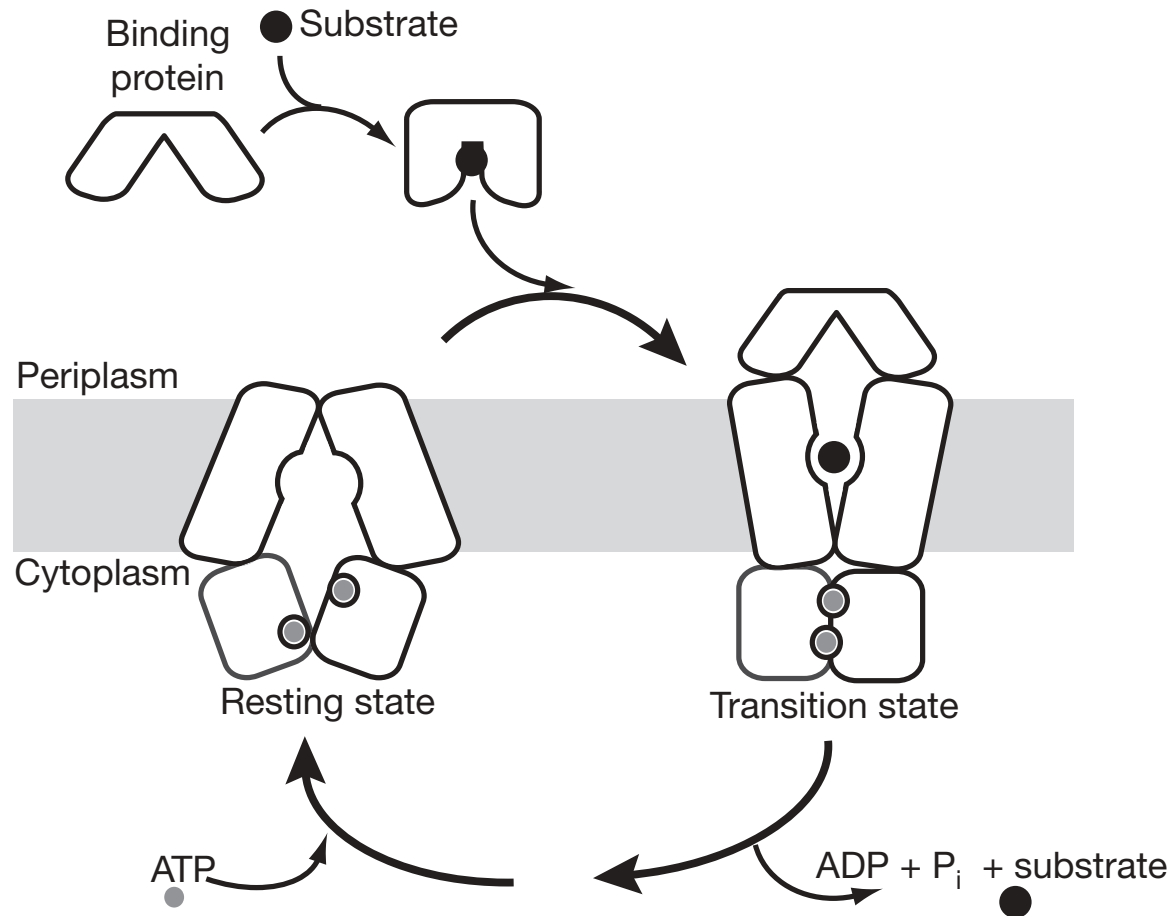
(Olham et al, Nature, 2007)



(Hollenstein et al, Nature, 2007)



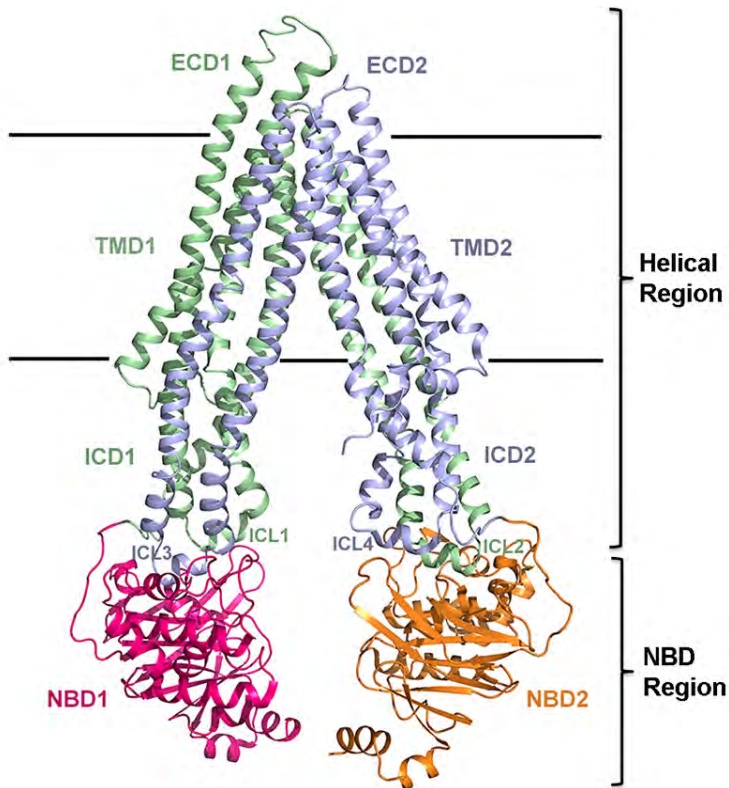
# Maltose transporter: a bacterial importer



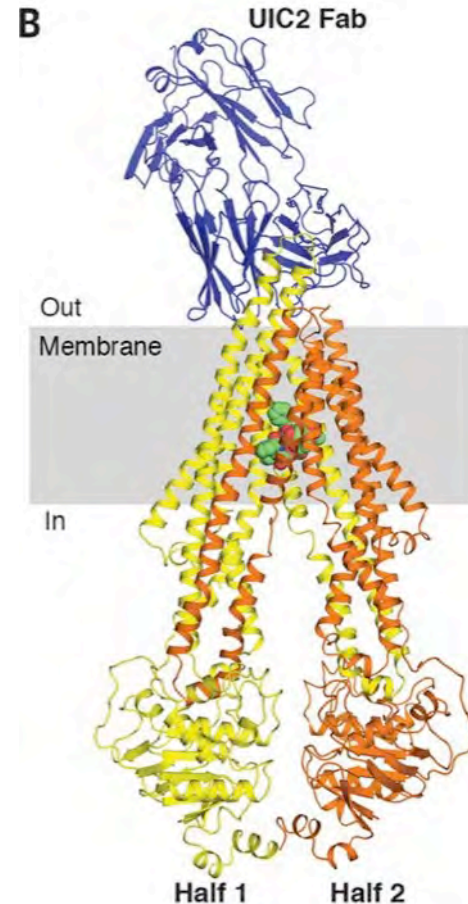
*(Olham et al, Nature, 2007)*



# P-glycoprotein: a multidrug-resistance efflux pump

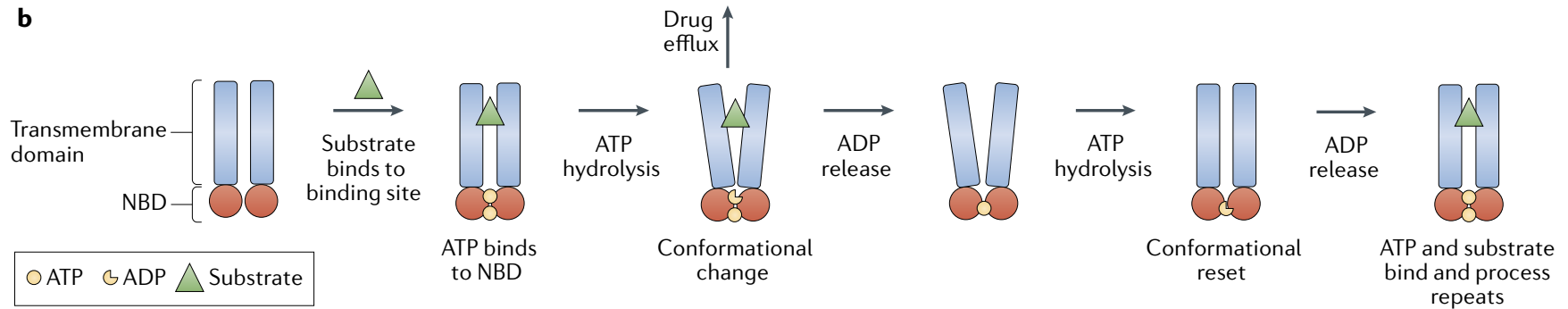


(Esser et al, JBC, 2017)



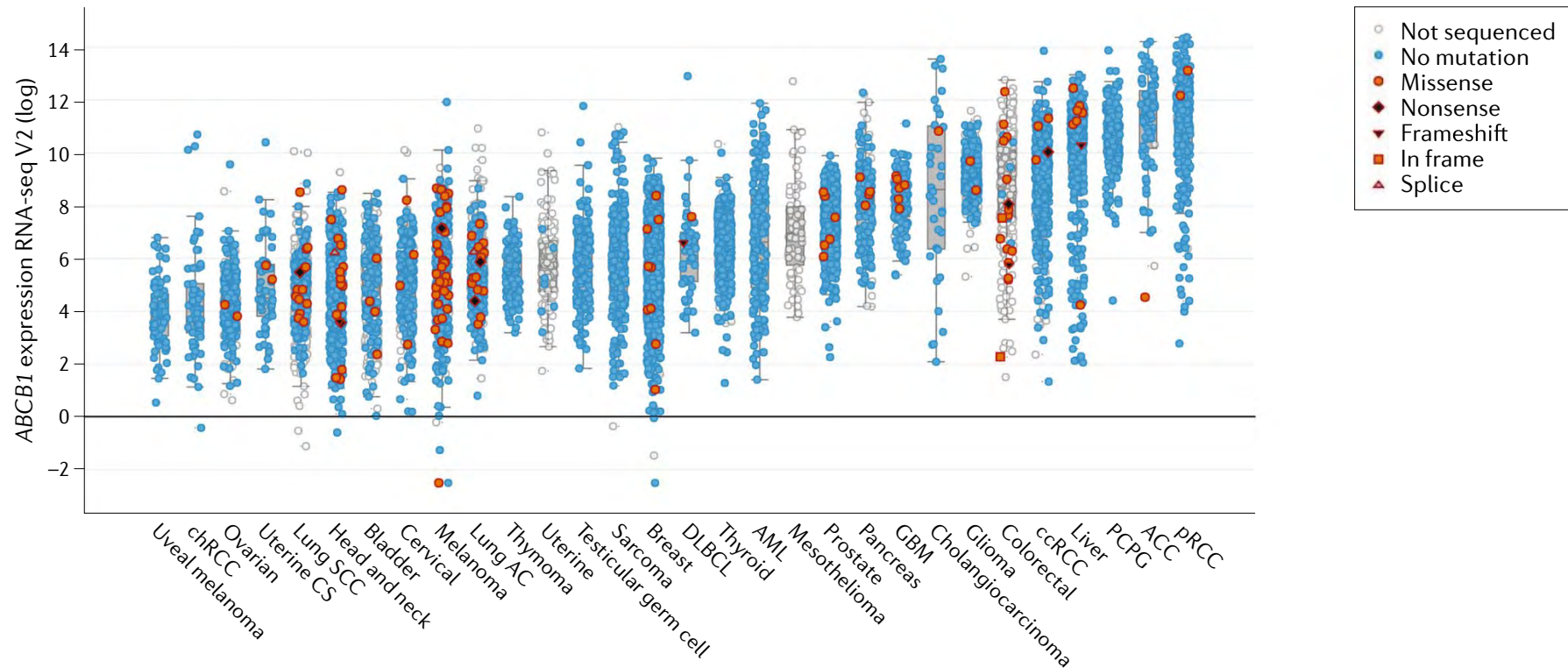
(Alam et al, Science, 2019)

# P-glycoprotein: a multidrug-resistance efflux pump



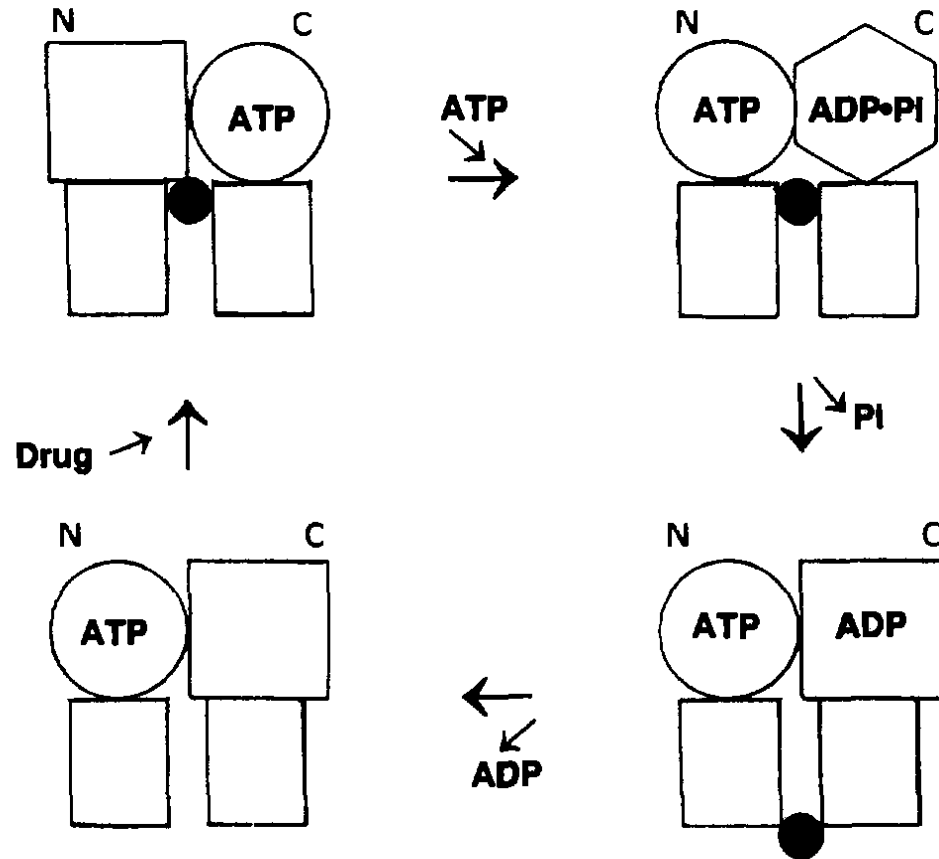
(Robey et al, Nat Rev Cancer, 2018)

# P-glycoprotein: high expression in cancer cells (cBioPortal + TCGA)

**a**

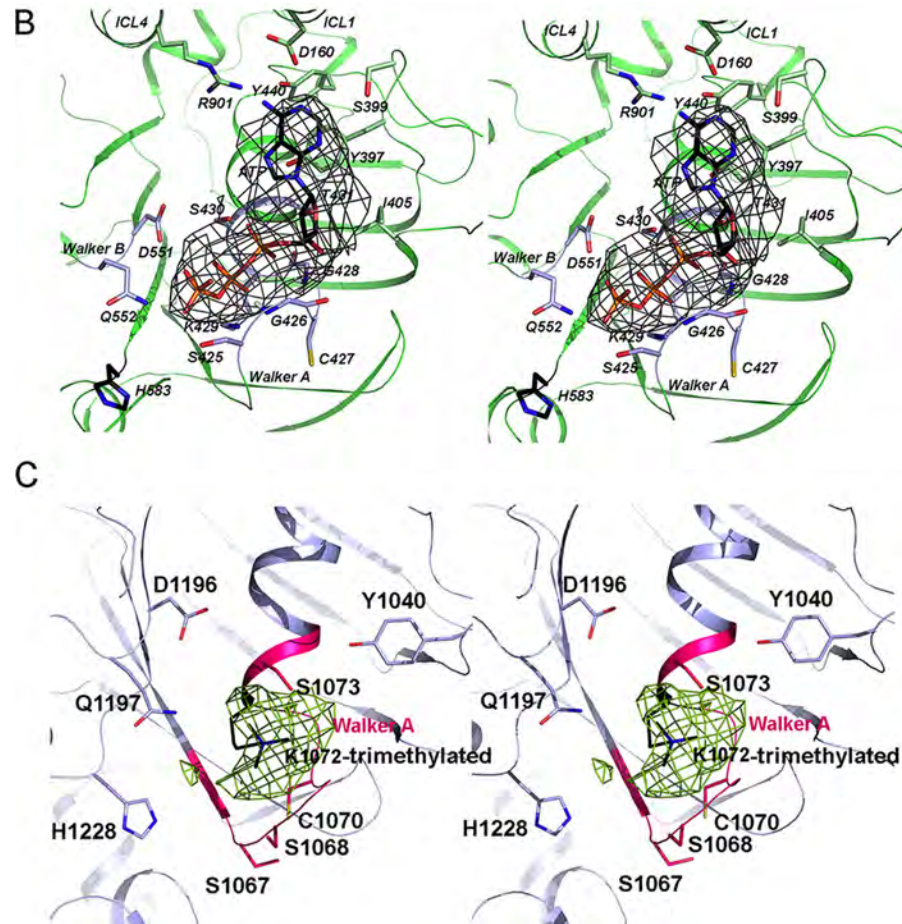
(Robey et al, Nat Rev Cancer, 2018)

# ABC: alternating catalytic cycle



(Senior et al, FEBS Lett, 1995)

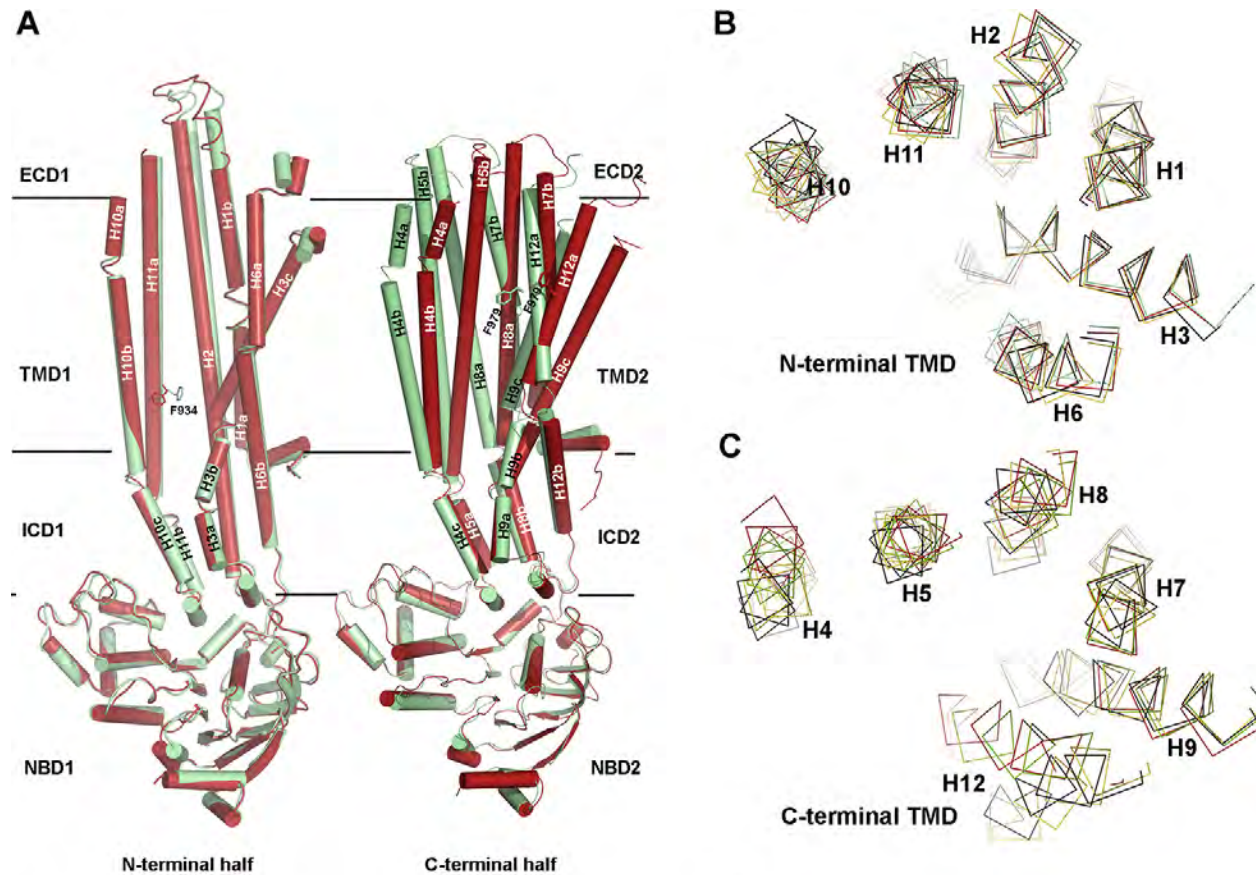
# P-glycoprotein: asymmetric ATP binding



(Esser et al, JBC, 2017)

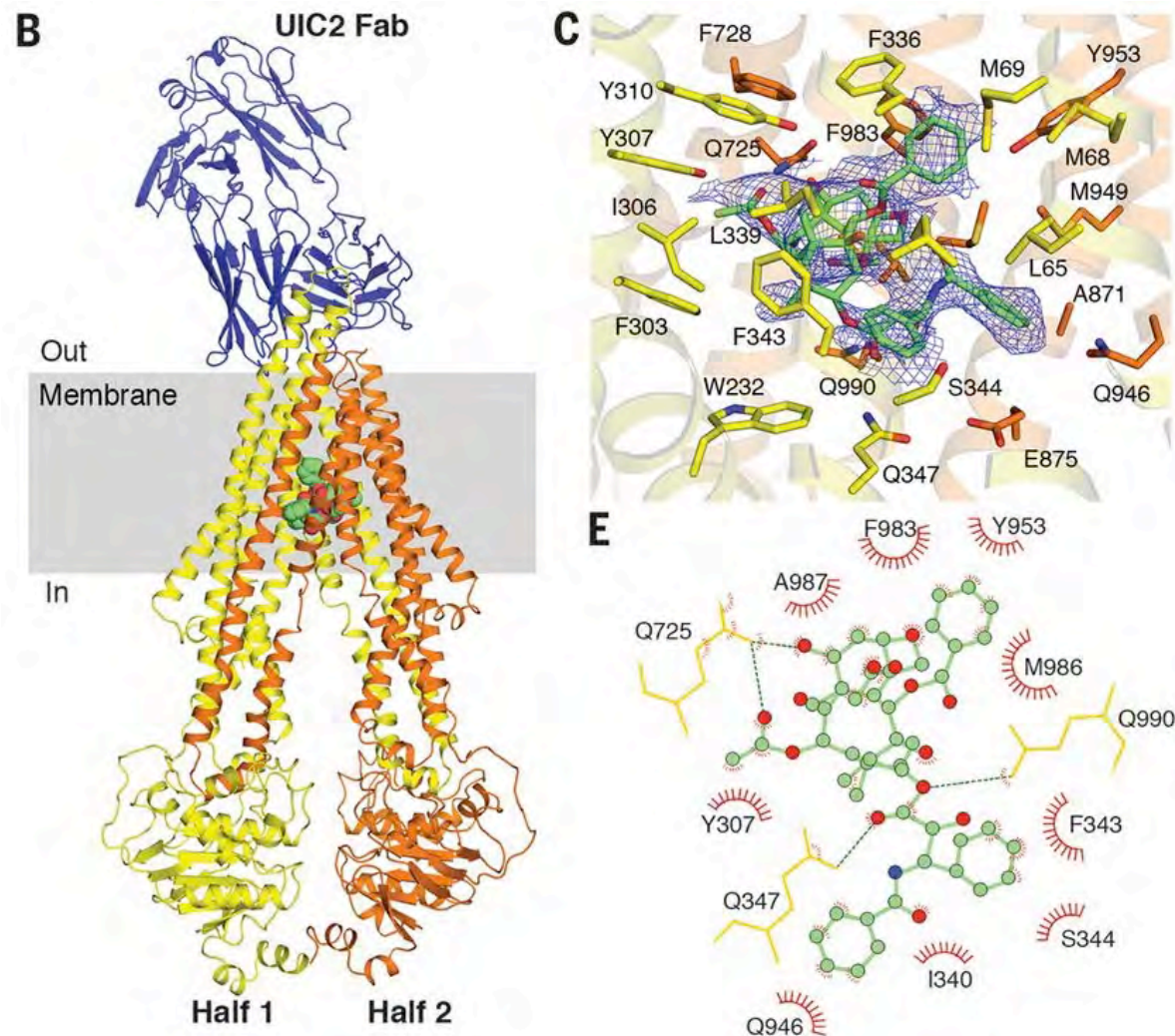


# P-glycoprotein: flexible TMD

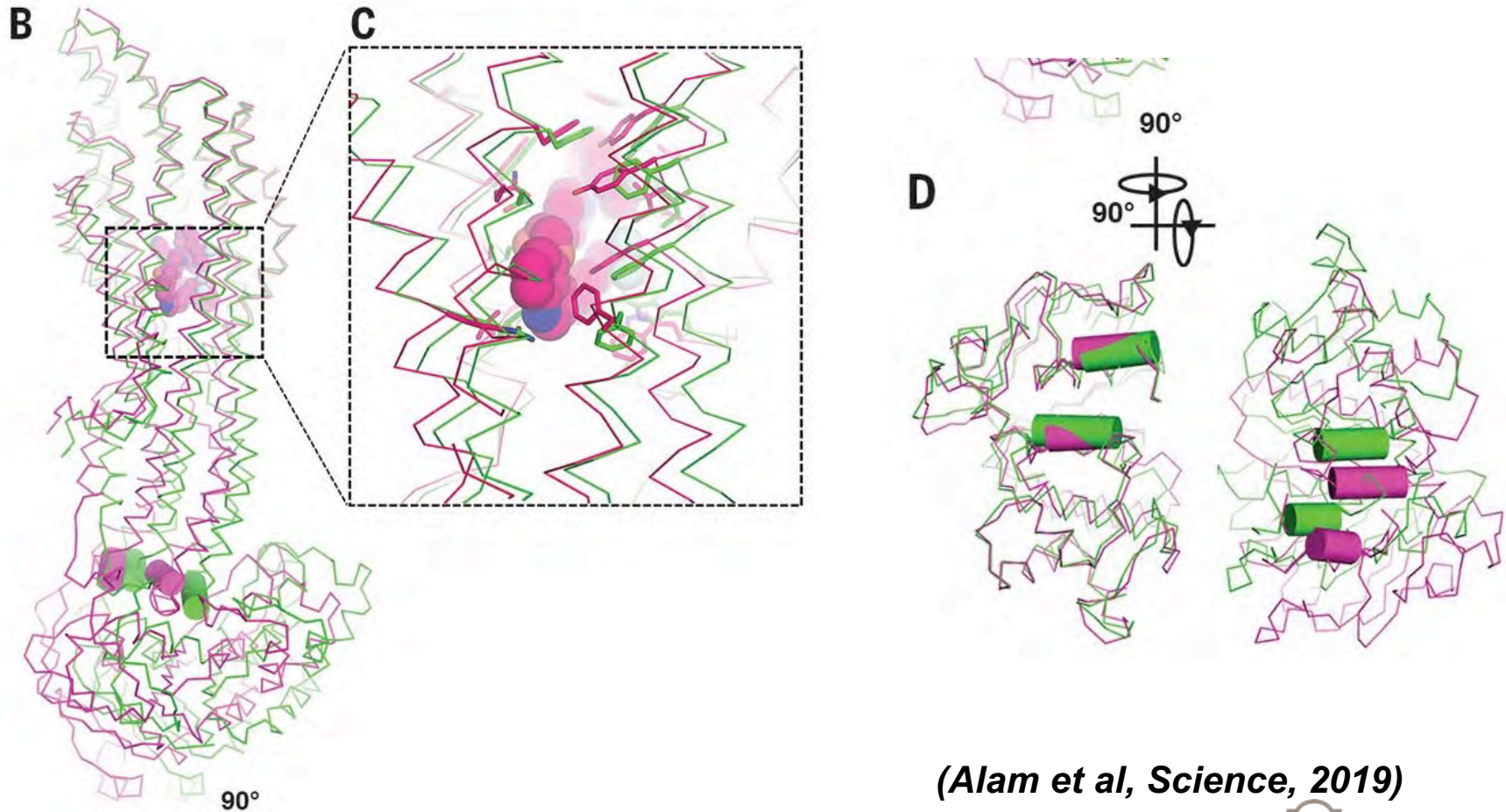


(Esser et al, JBC, 2017)

# P-glycoprotein: inhibitor v.s. drug (Taxol)



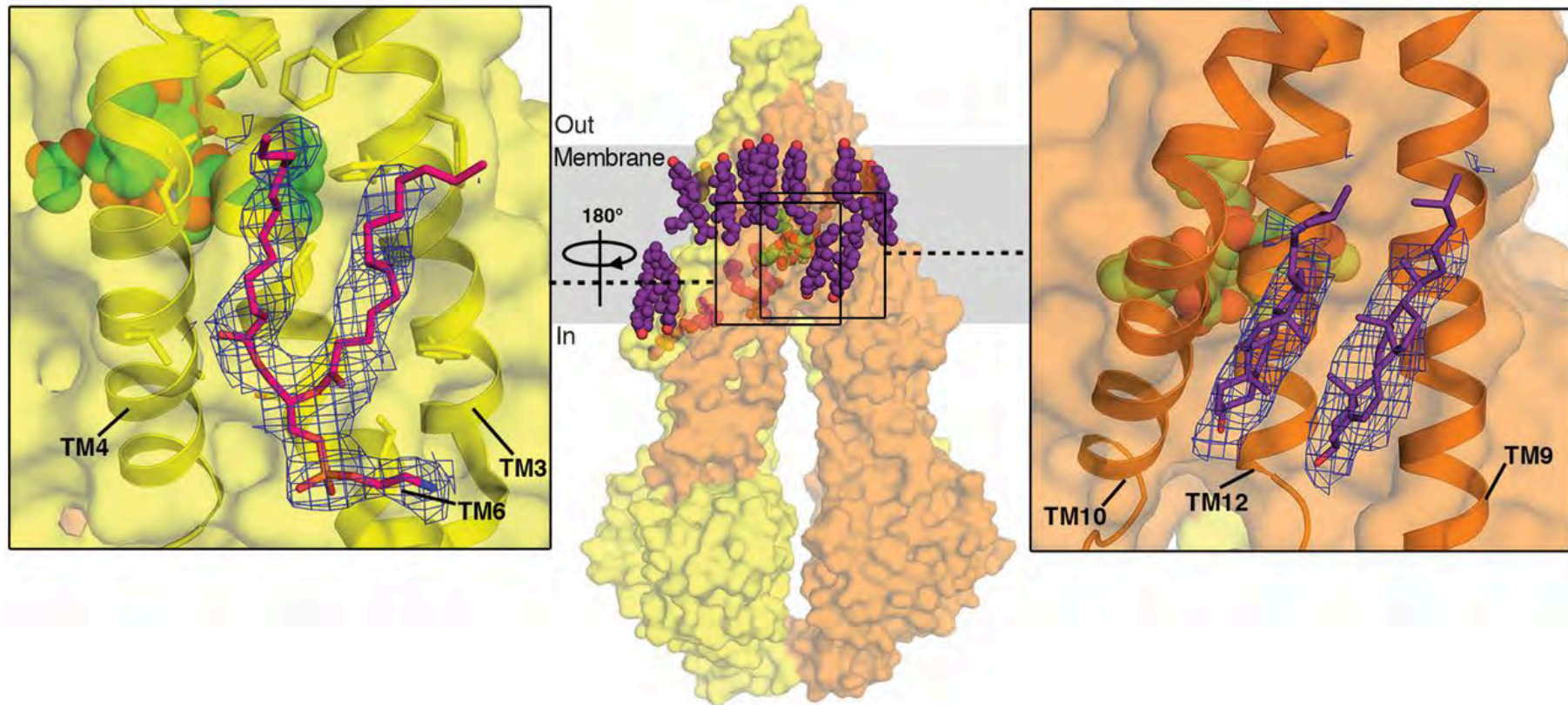
# P-glycoprotein: transport substrates affect ATP usage



(Alam et al, Science, 2019)

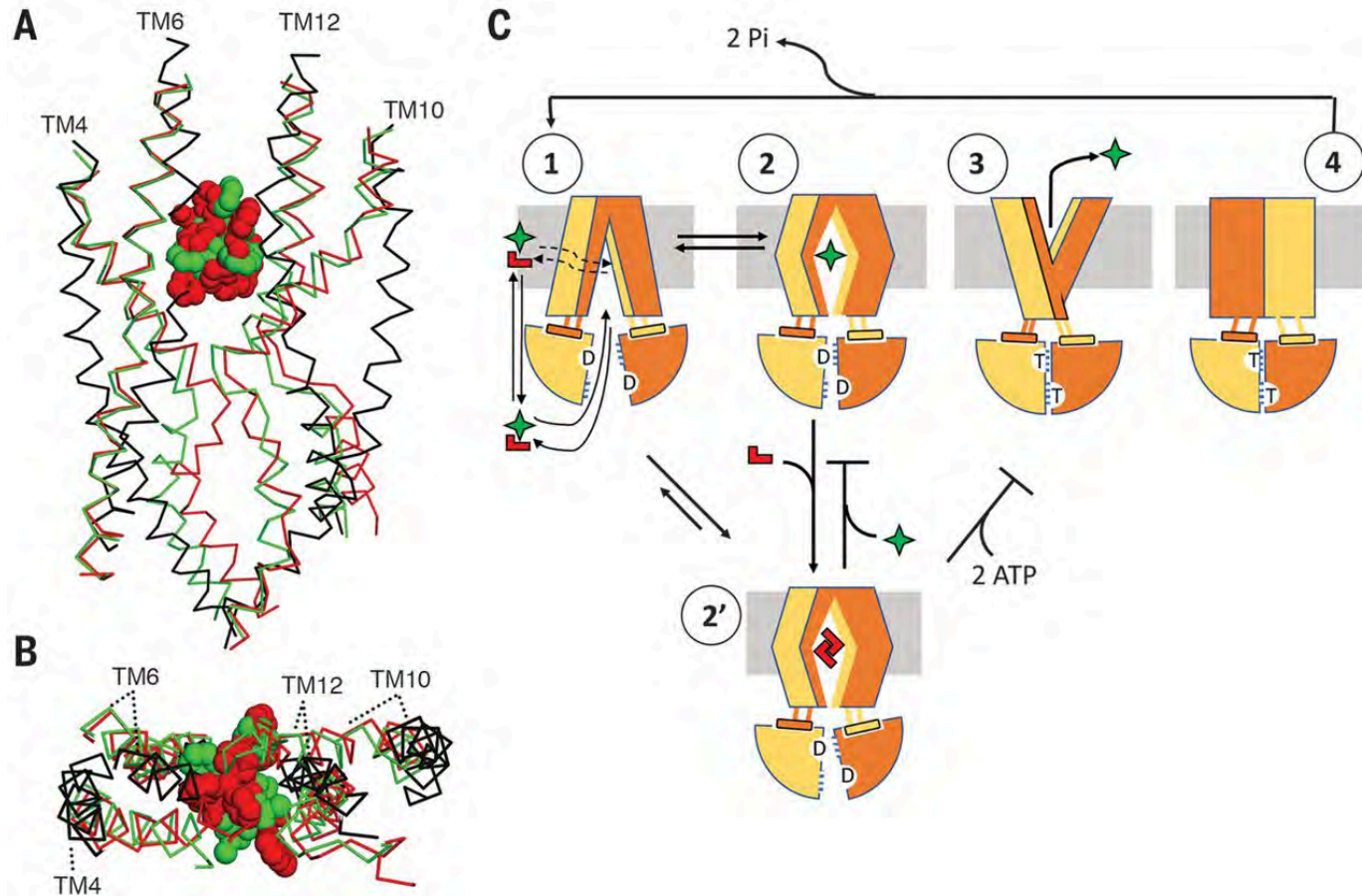


# P-glycoprotein: lipid-protein interaction



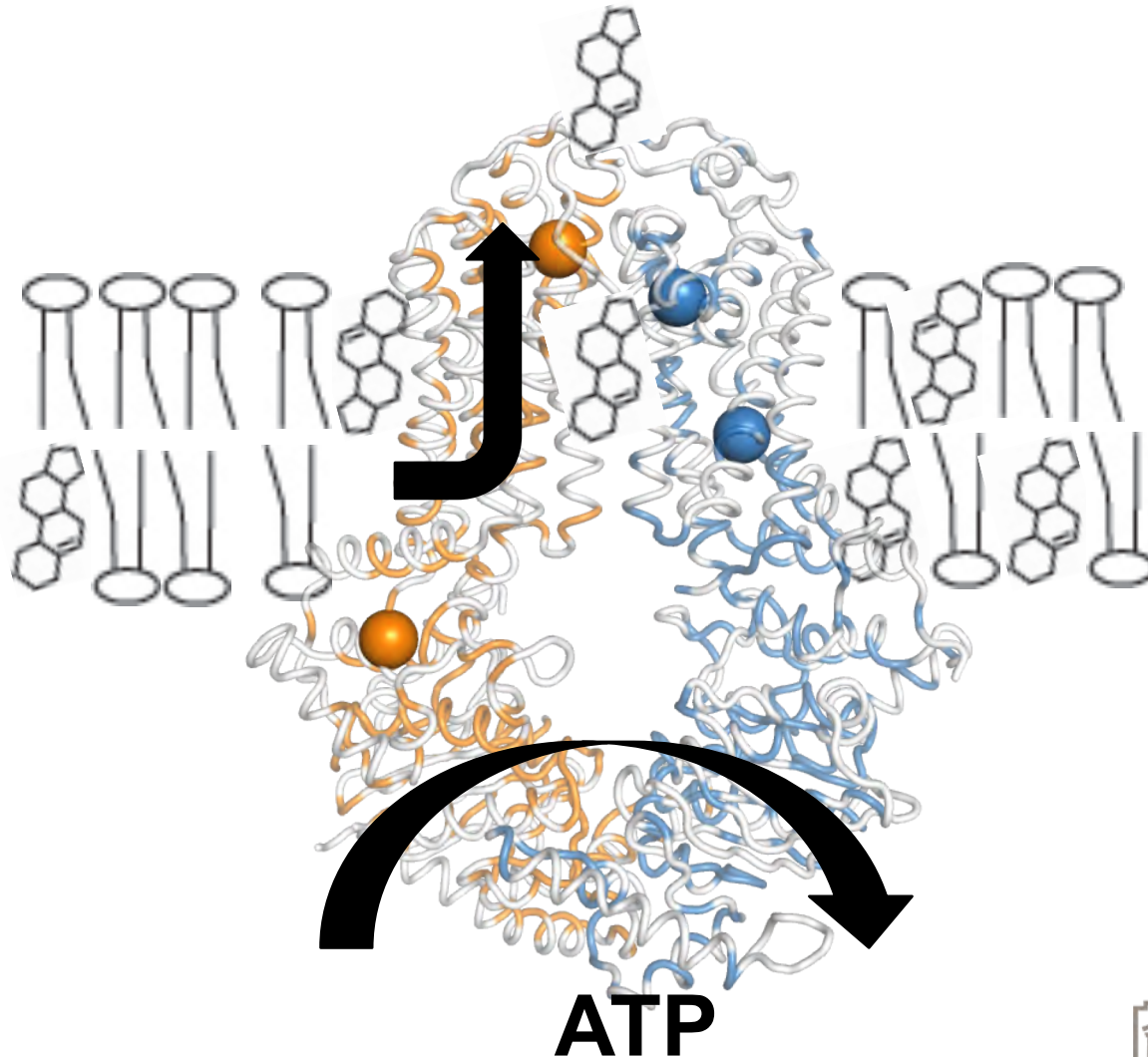
(Alam et al, Science, 2019)

# P-glycoprotein: multidrug resistance?



(Alam et al, Science, 2019)

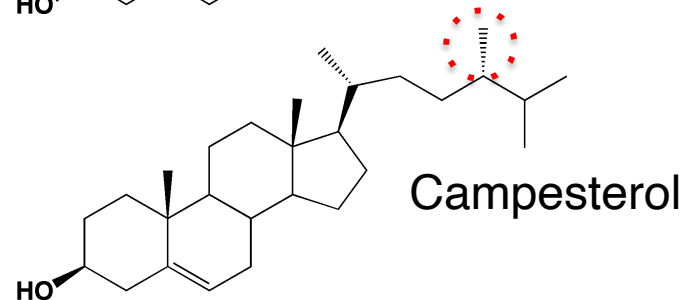
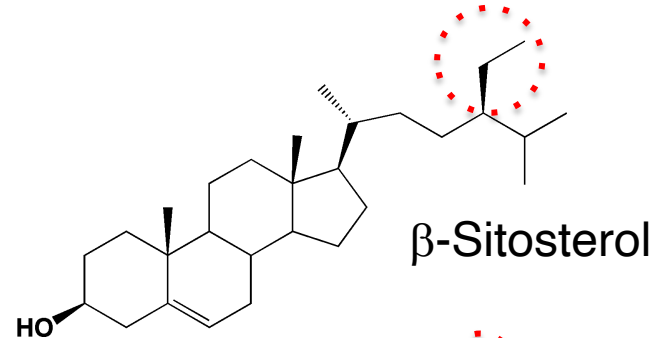
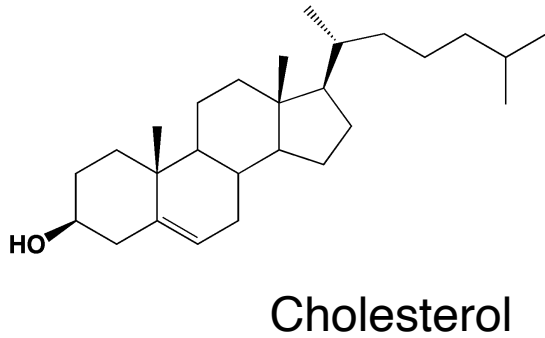
# ABCG5/G8: a sterol/cholesterol efflux pump



# Dietary Sterols

## Animal (60%)

## Plant (40%)

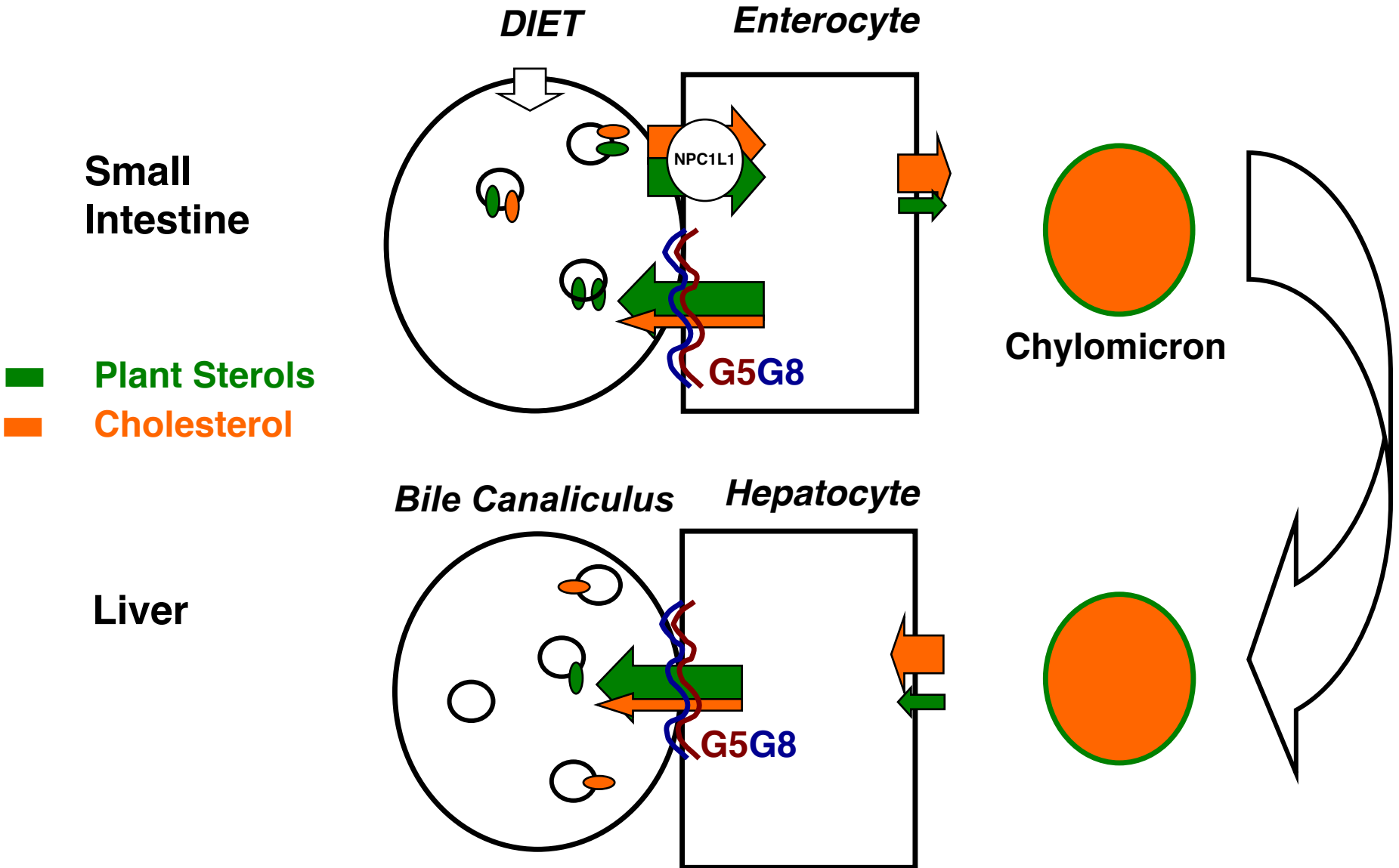


ABSORPTION:  $\sim 50\%$

$< 5\%$



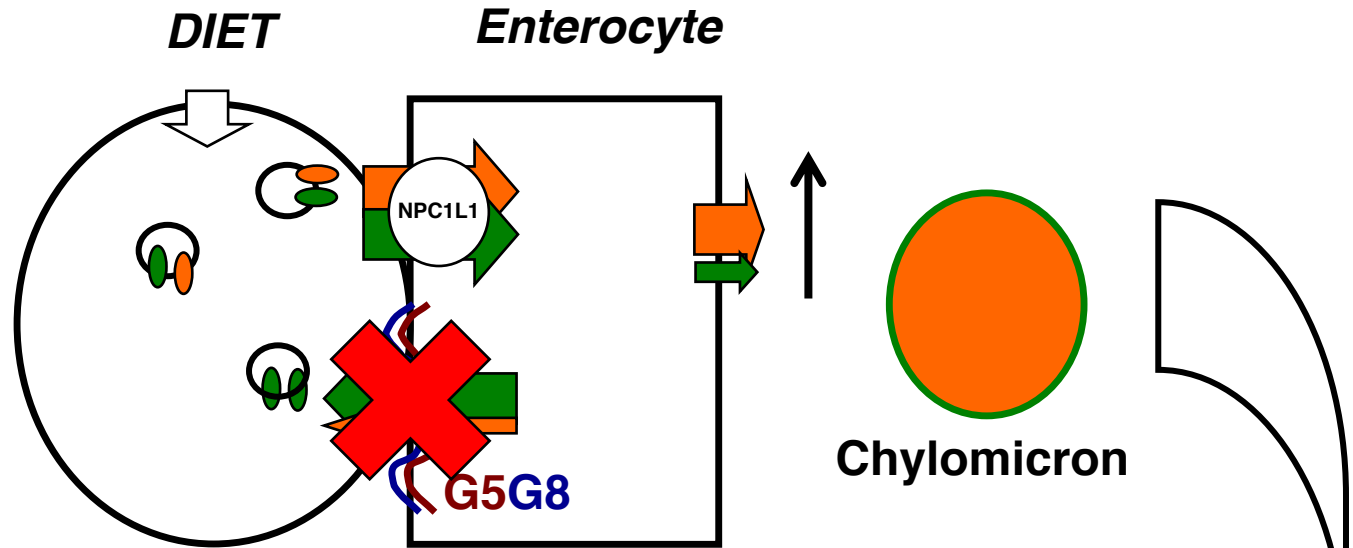
# ABCG5/G8 promotes biliary and intestinal sterol secretion (liver/small intestine specific).



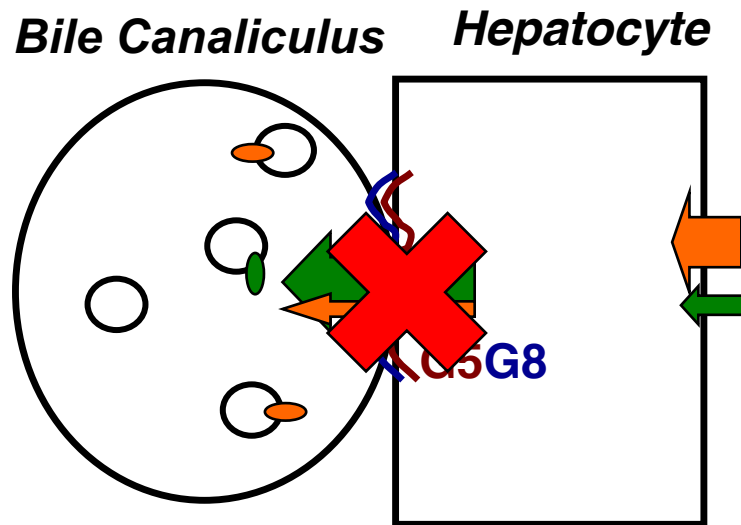
# G5G8 protects against plant sterol accumulation. (Sitosterolemia)

■ Plant Sterols  
■ Cholesterol

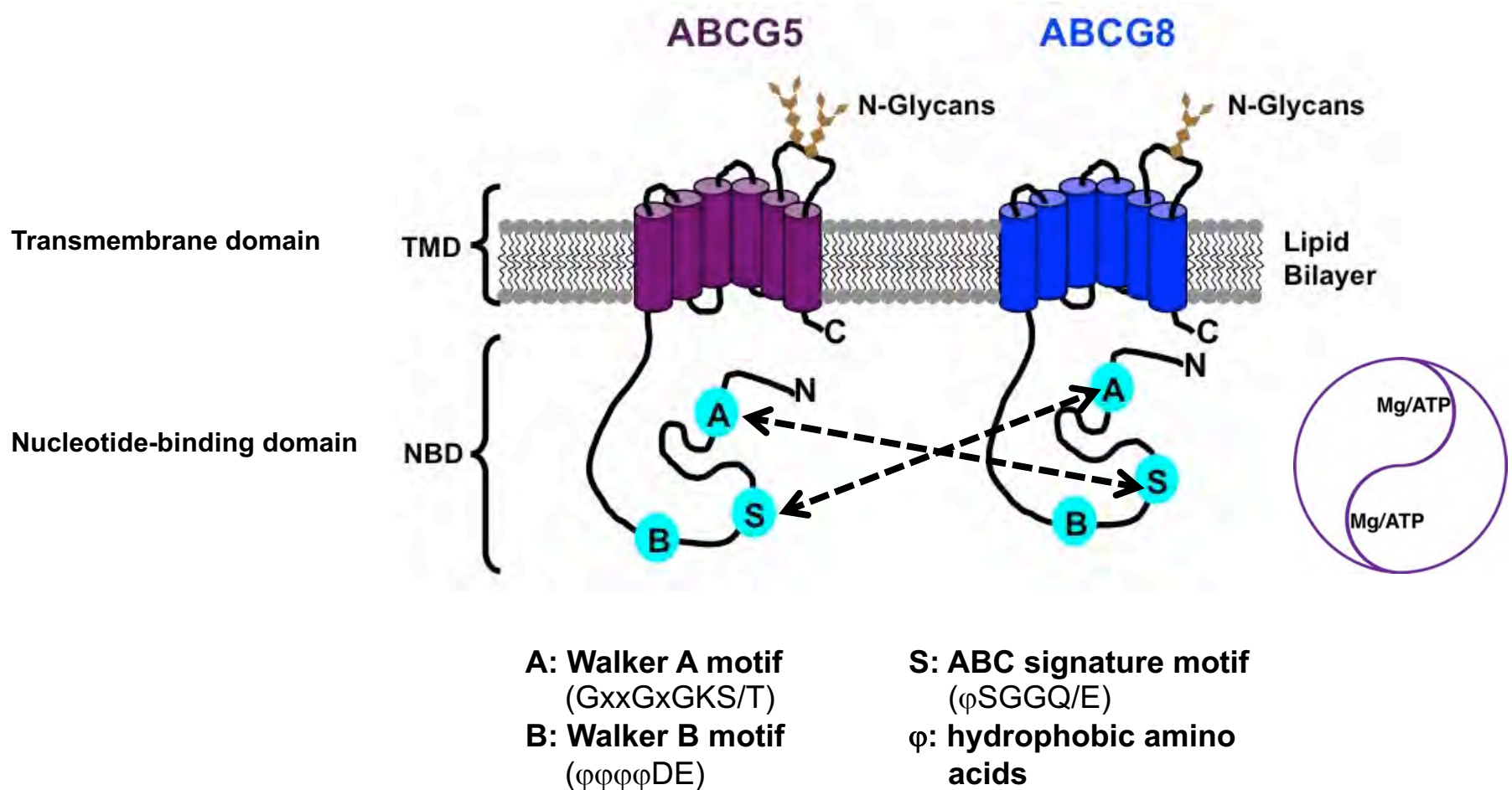
Small  
Intestine



Liver

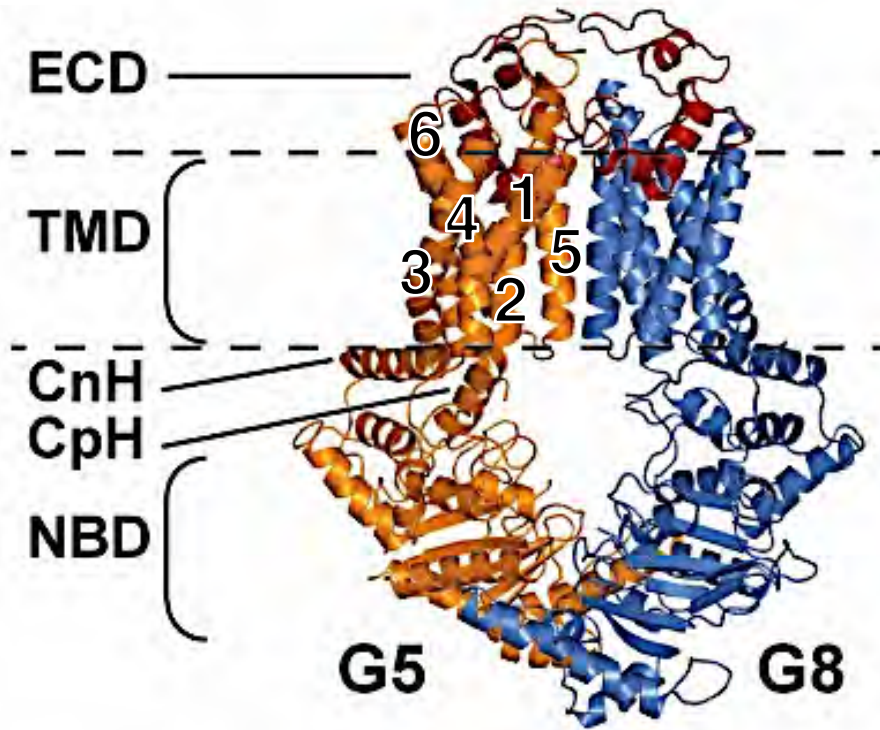


# ABCG5 and ABCG8 are half ABC transporters.



# ABCG5 and ABCG8 share high structural similarity.

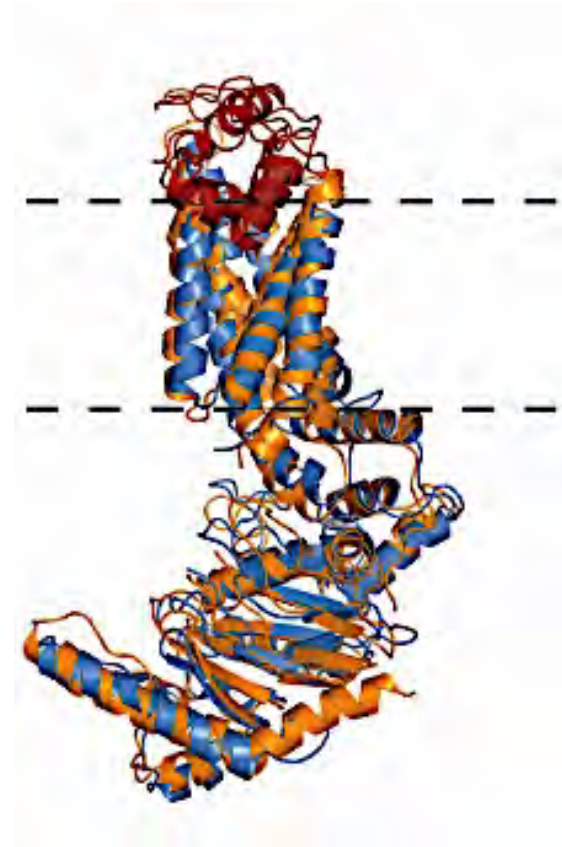
## Domain features



TMD: transmembrane domain  
NBD: nucleotide-binding domain

ECD: extracellular domain  
CnH: connecting helix  
CpH: coupling helix

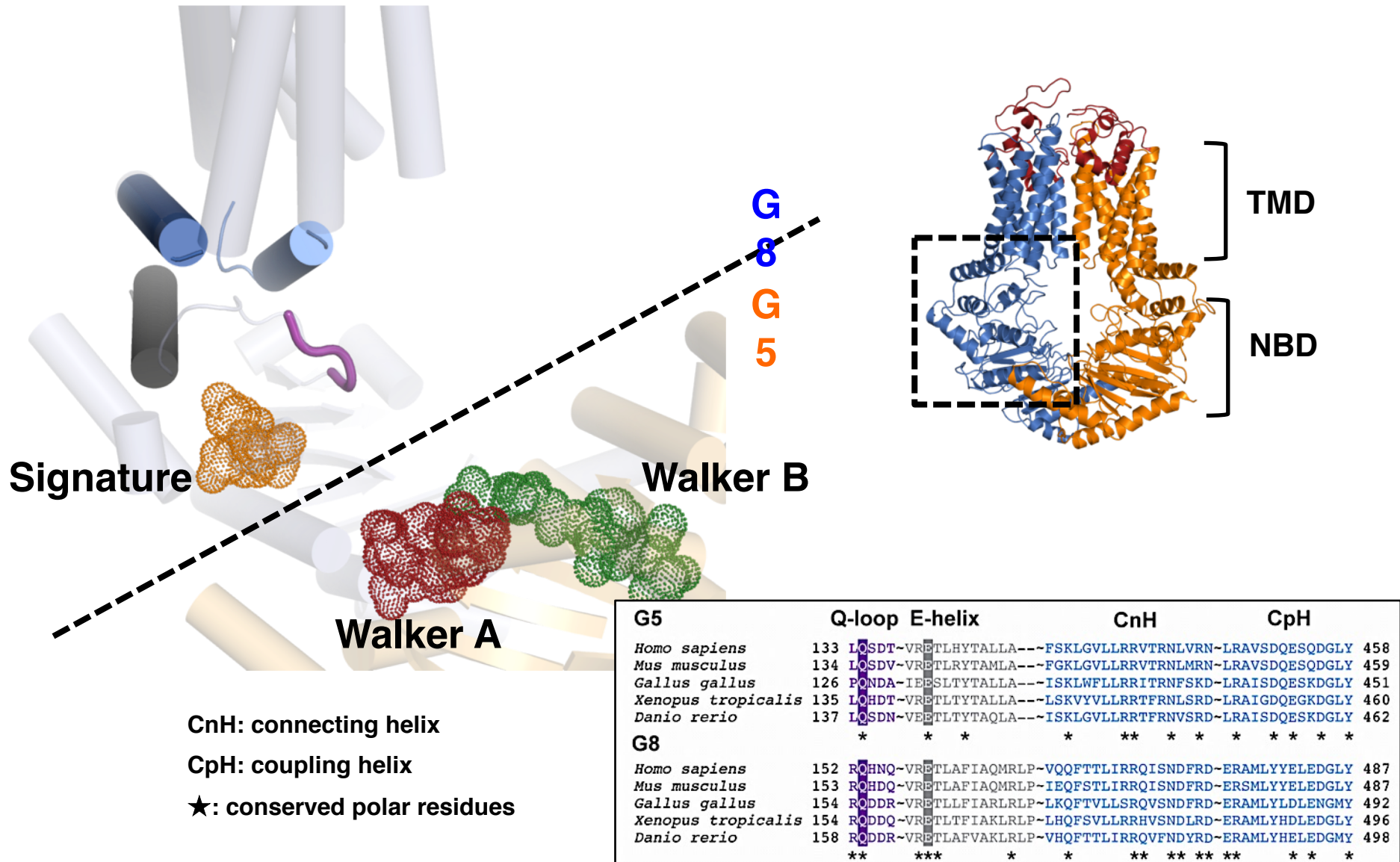
## Structural similarity:



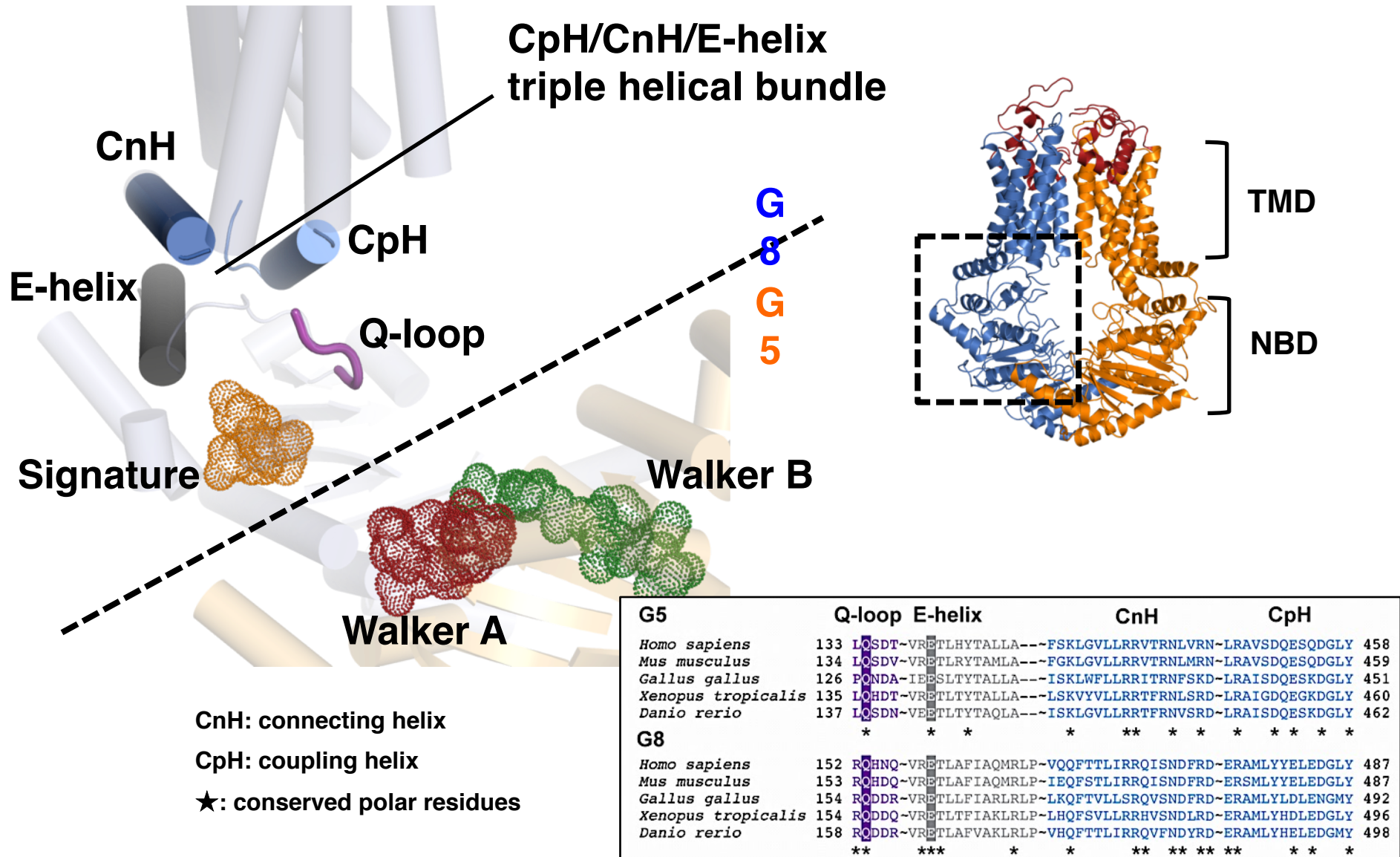
RMSD ( $C\alpha$ )  $\sim 2\text{\AA}$   
( $\sim 28\%$  sequence identity)



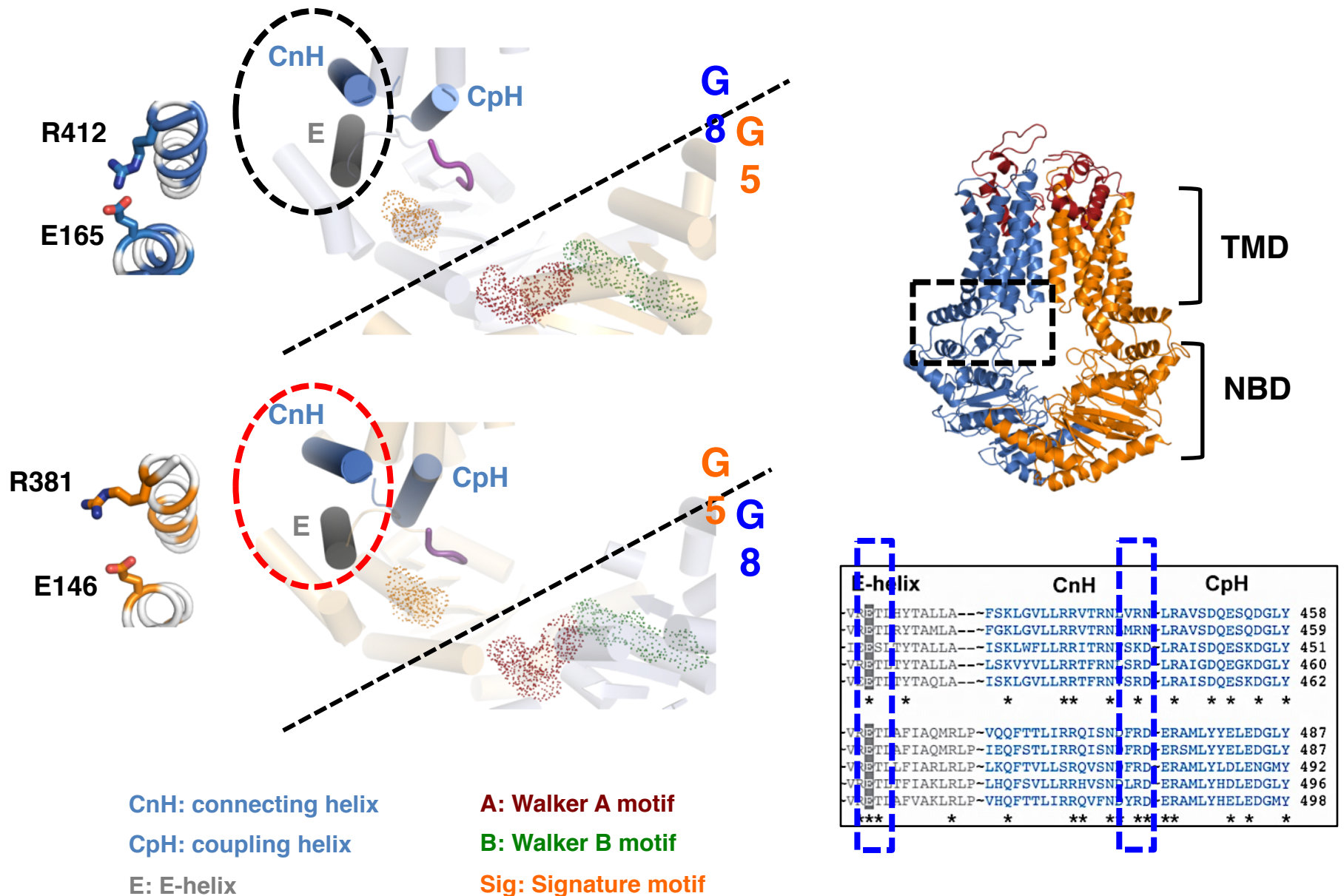
# Triple Helical Bundle: Connecting the ATP-Binding Cassette to the Transmembrane Domain



# Triple Helical Bundle: Connecting the ATP-Binding Cassette to the Transmembrane Domain

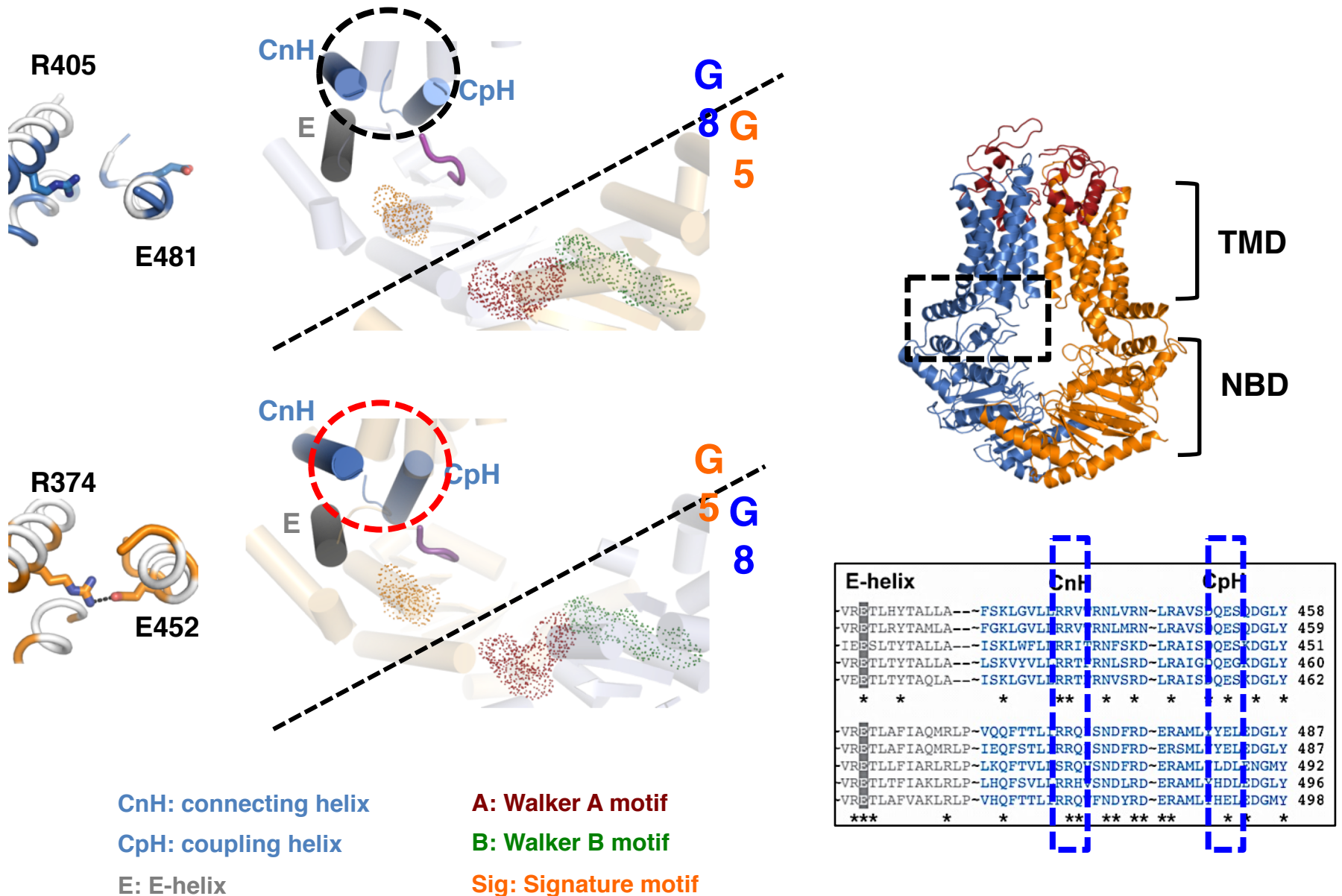


# Interactions within the Triple Helical Bundle

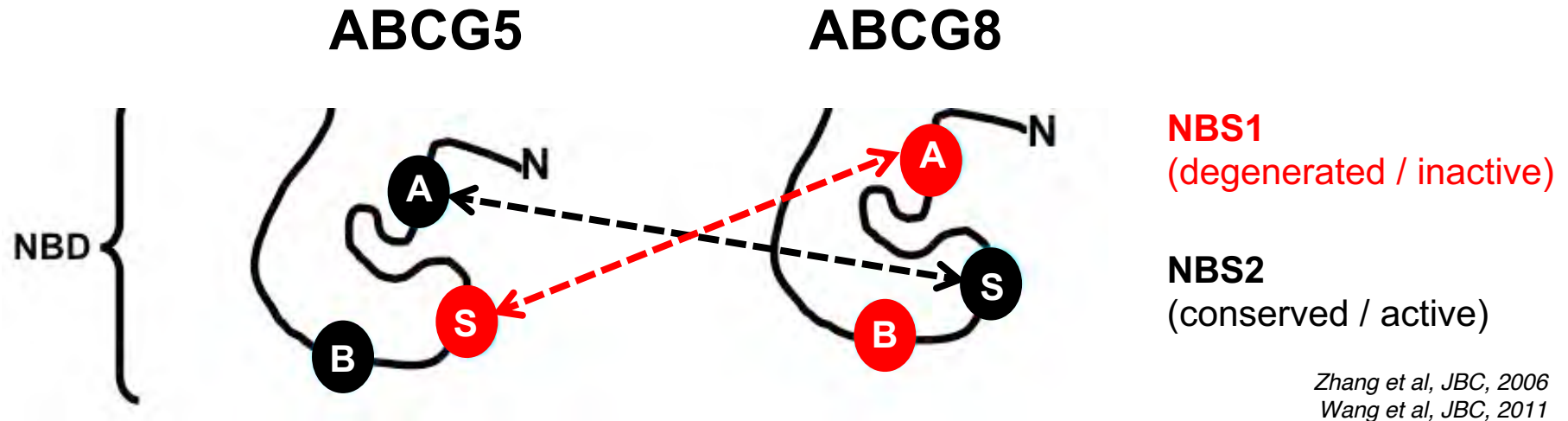




# Interactions within the Triple Helical Bundle



# The ATPase in G5G8 is catalytically asymmetric.



## Consensus Sequence

GxxGxGKS/T

φSGGQ/E

φφφφDE

(φ: hydrophobic amino acids)

**A:** Walker A motif

GSSGSGKT

GSSGCG**RA**

**S:** Signature motif

IST**T**GE

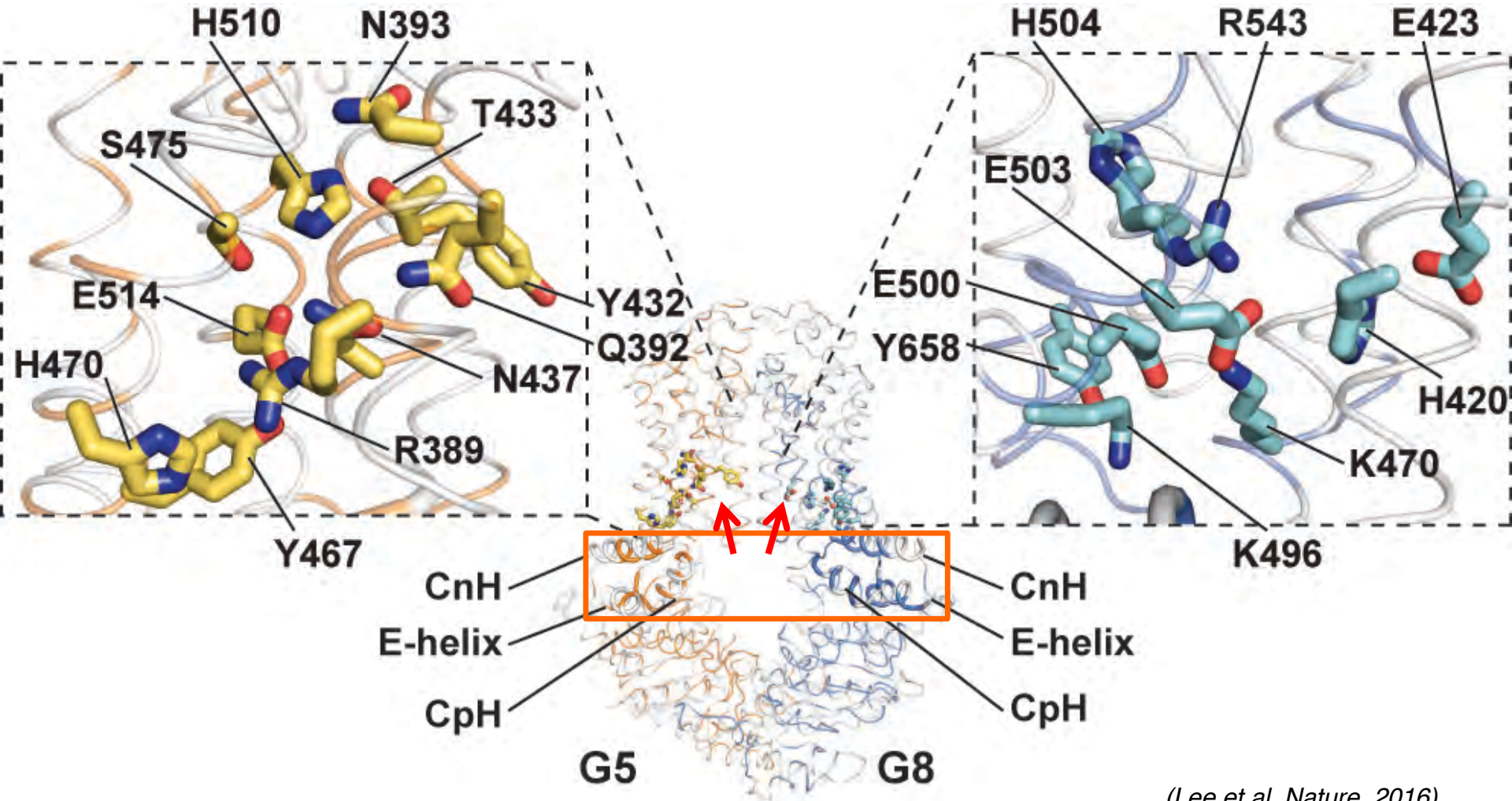
LSGGE

**B:** Walker B motif

VMLFDE

ILILDE

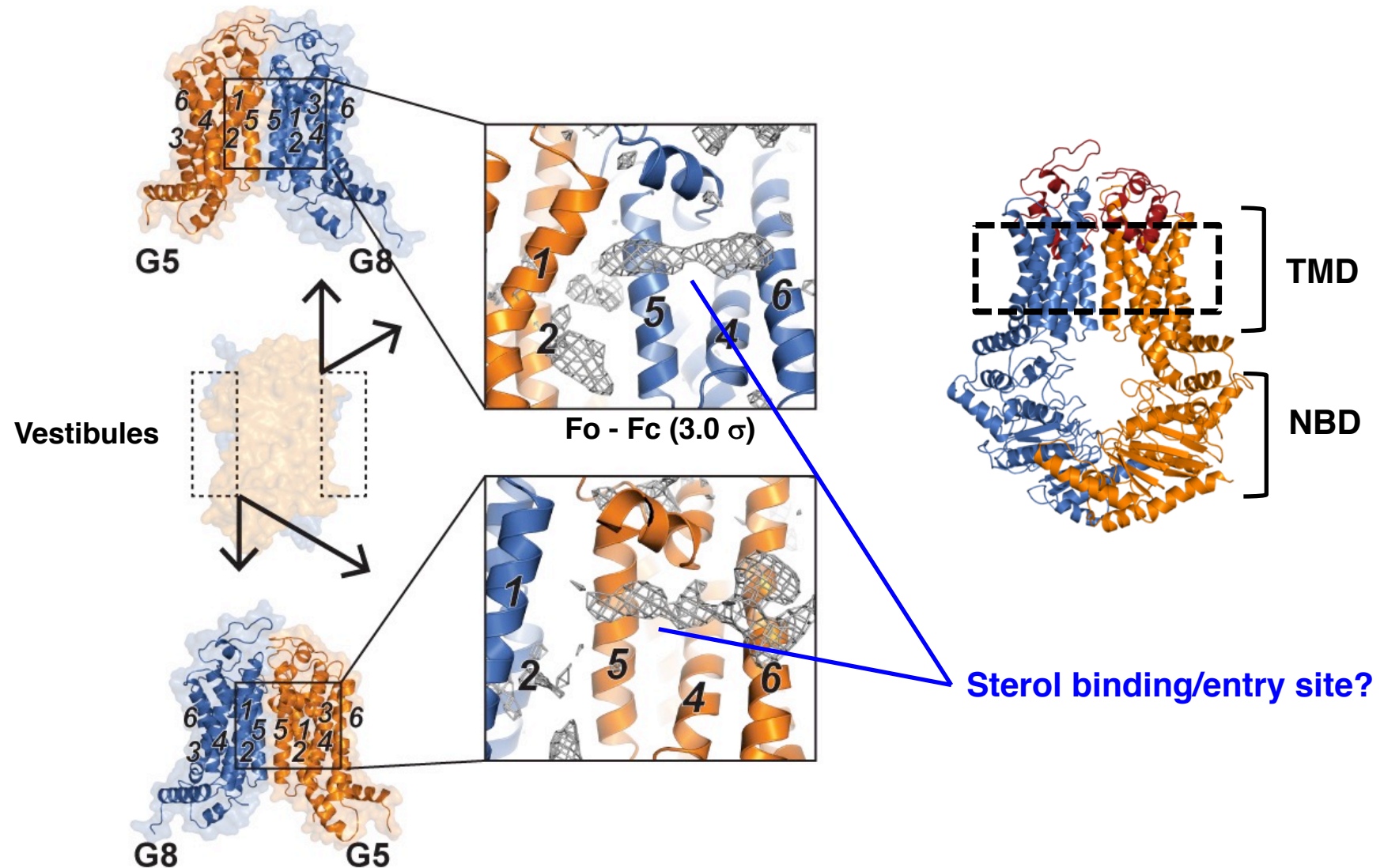
# The TMD polar relay connects the triple helical bundle to the TMD.





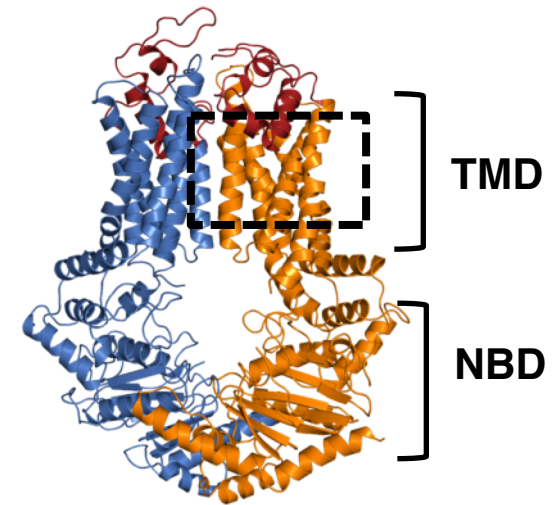
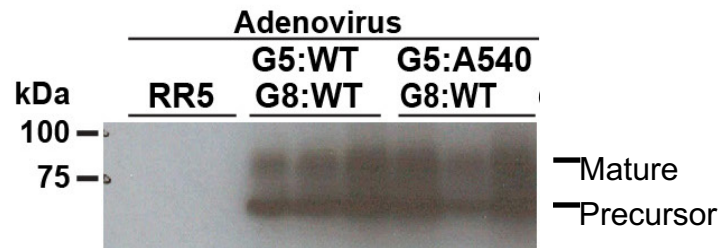
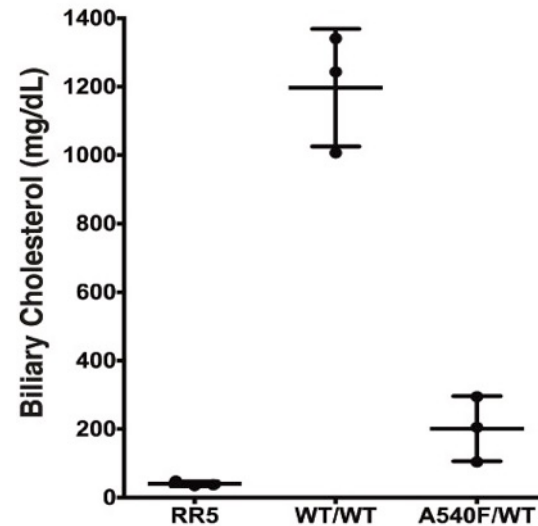
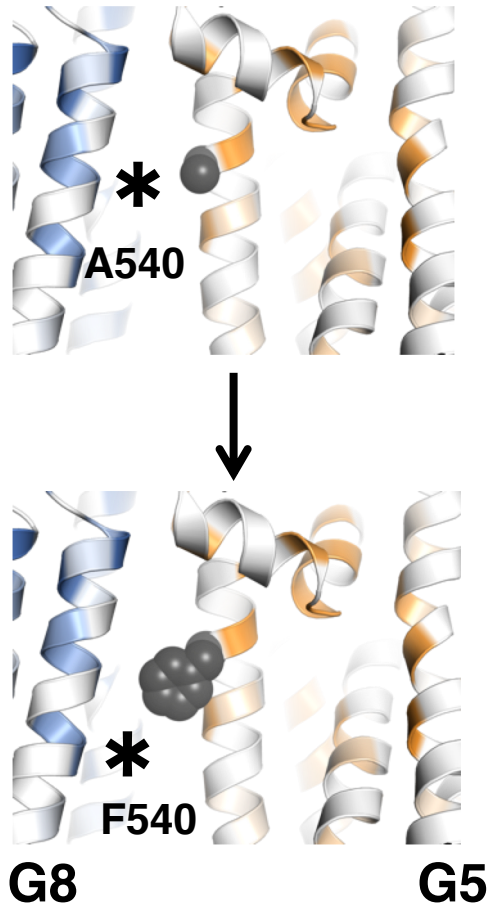
# How do sterols move across the lipid-bilayer membranes on the TMD?

## Vestibules at the TMD-membrane interface





# How do sterols move across the lipid-bilayer membranes on the TMD?

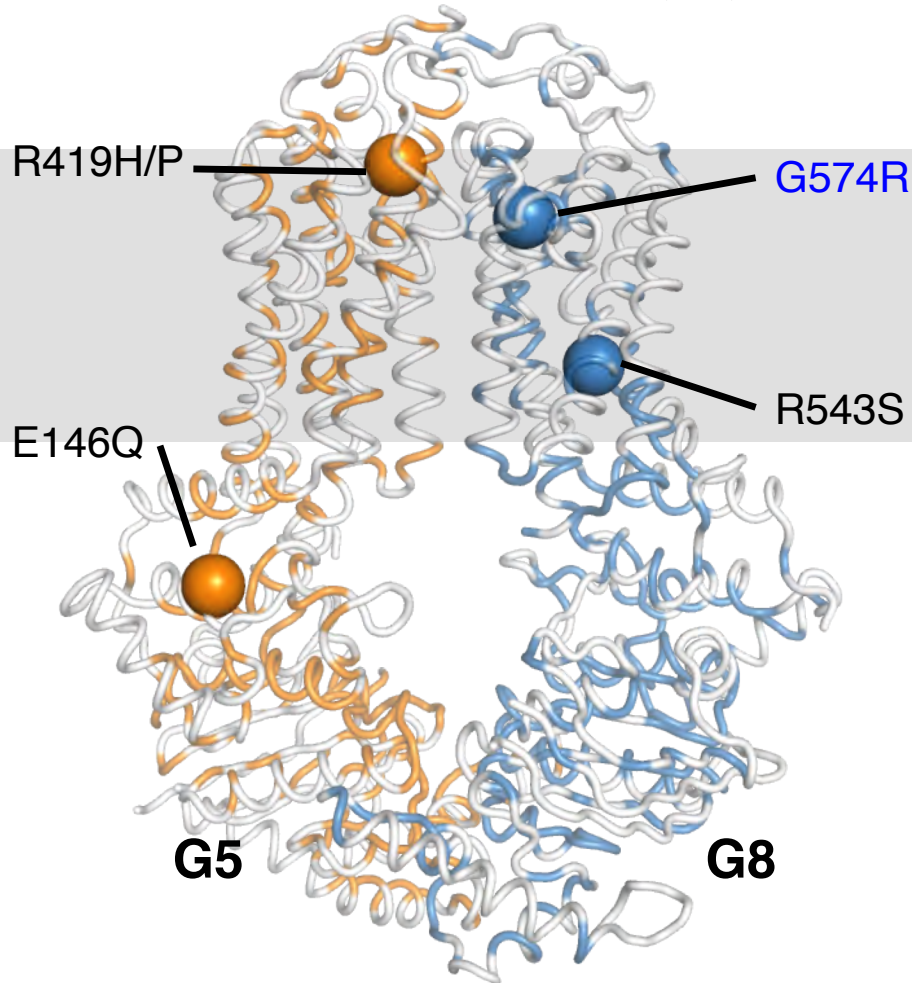


(Lee et al, Nature, 2016)

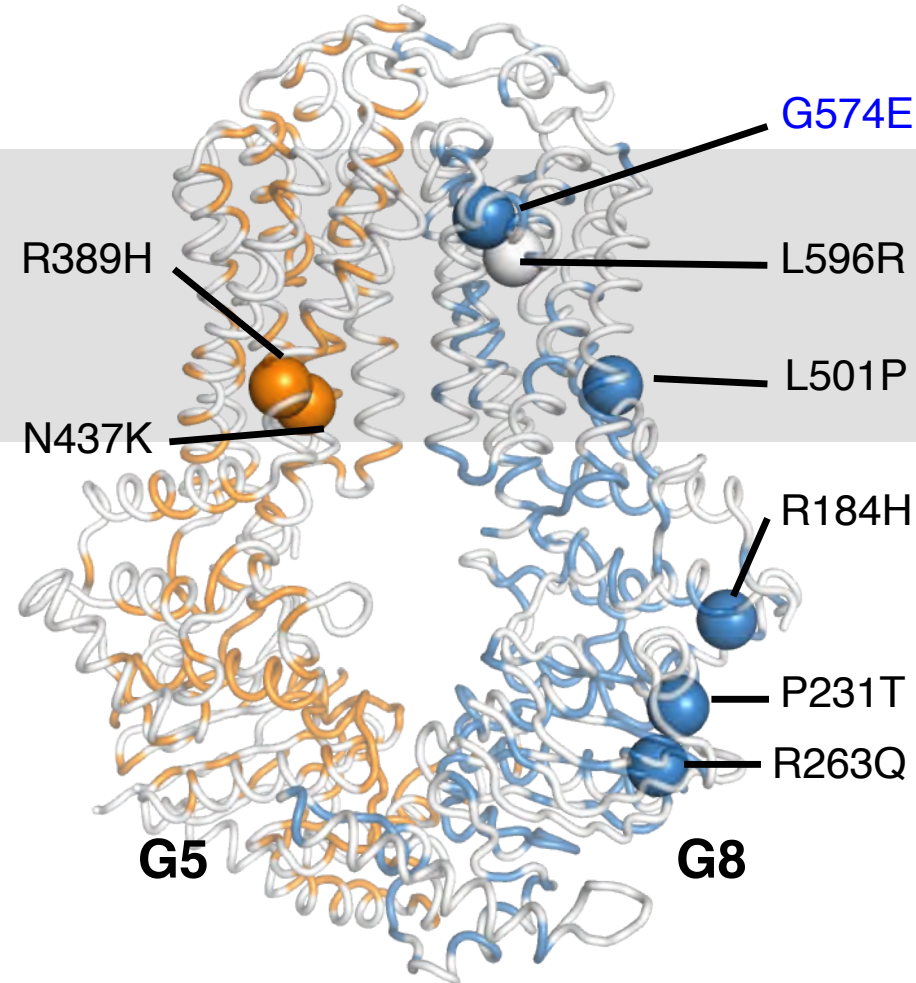
# Location of the residues with the disease-causing missense mutations of **sitosterolemia**.

## ER-escape missense mutations

*Graf et al, JBC, 2004*



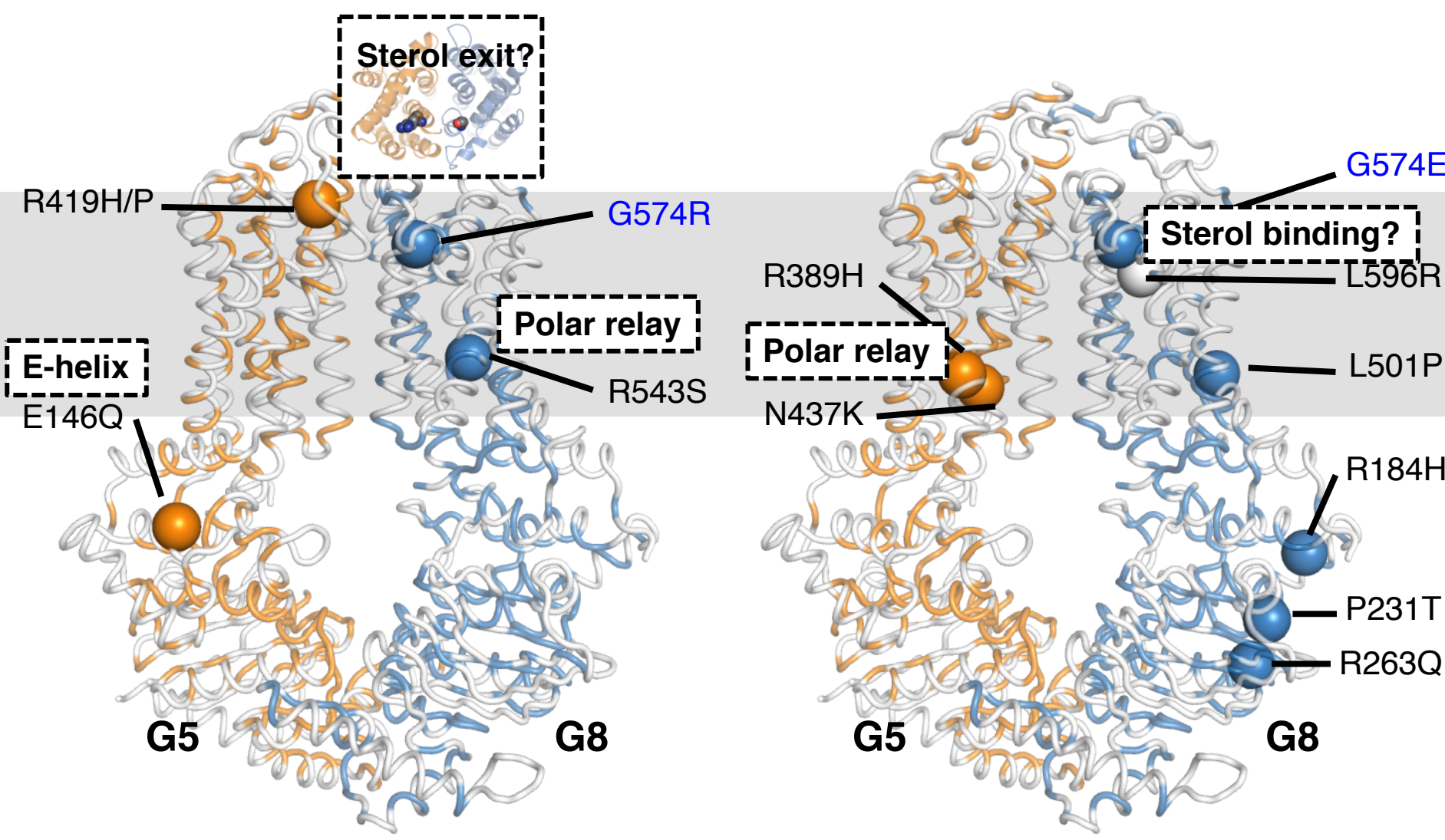
## Non-ER-escape missense mutations



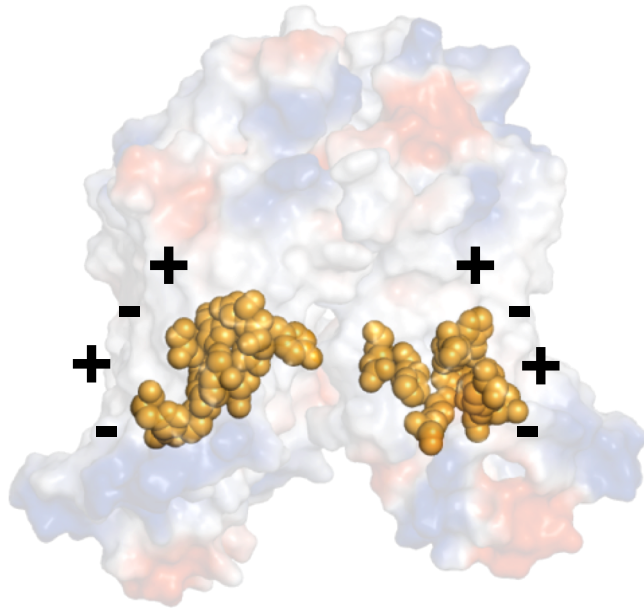
**Color:** conserved (multiple sequence alignment (MSA) value  $\geq 7$ )

**White:** less/non-conserved (MSA  $< 7$ )

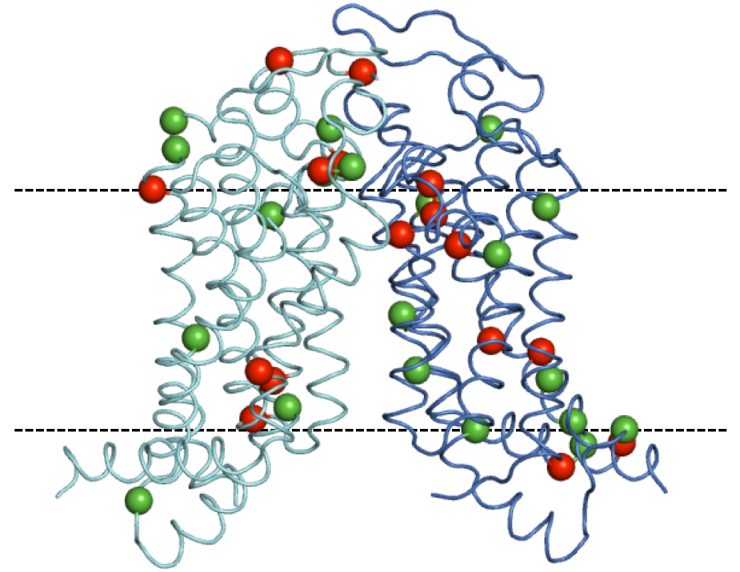
# Disease-causing mutations cluster in the conserved functional domains in G5G8.



# Transmembrane Domain of ABC Cholesterol Transporters: a Pathogenic Hot Spot



**Polar relay**



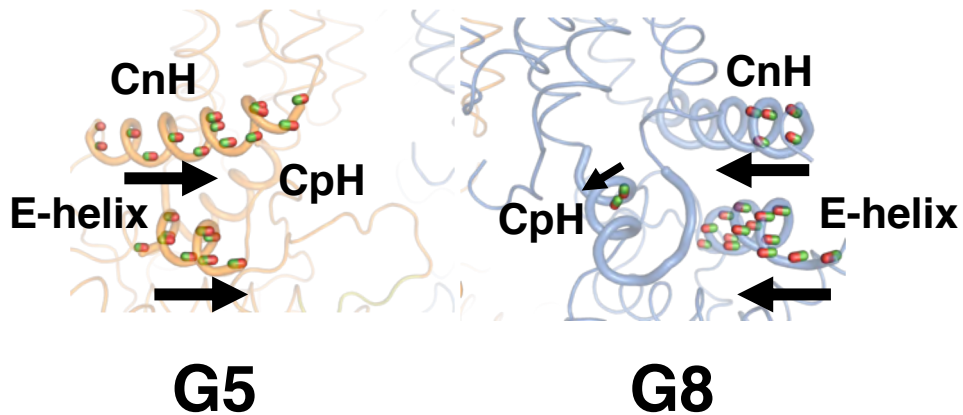
**Pathogenic residues:  
G5G8 (red), A1 (green)**



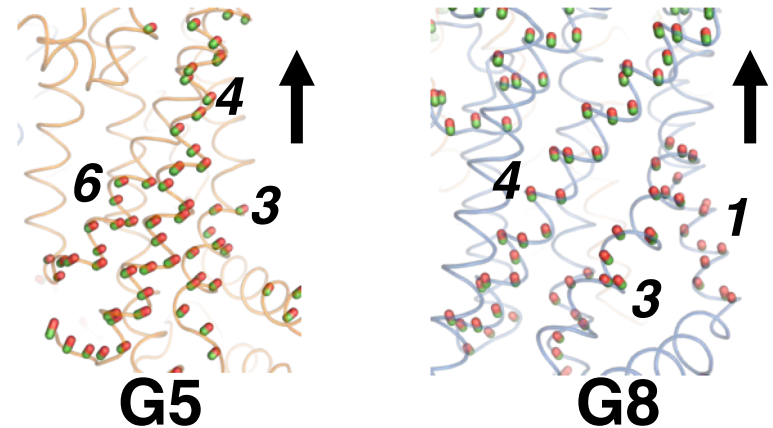
# G5G8-Mediated Sterol Transport

## Molecular Dynamics Simulation

### Inward movement (CpH/CnH/E-helix bundle)

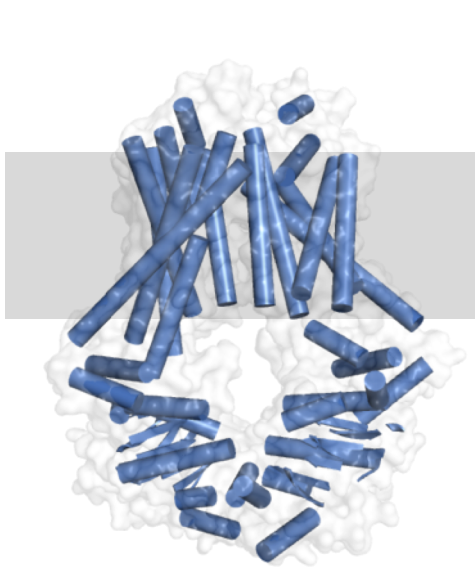


### Upward movement (TM helices)

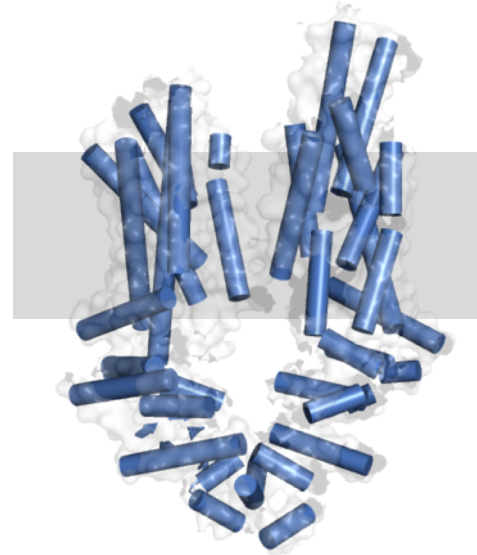
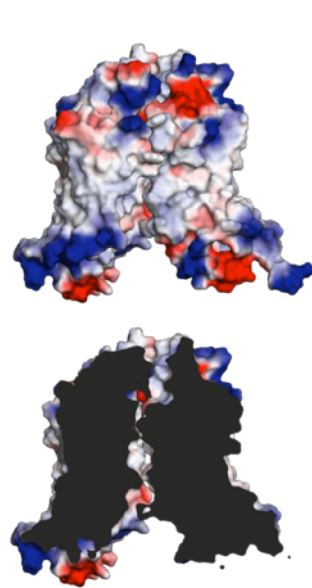




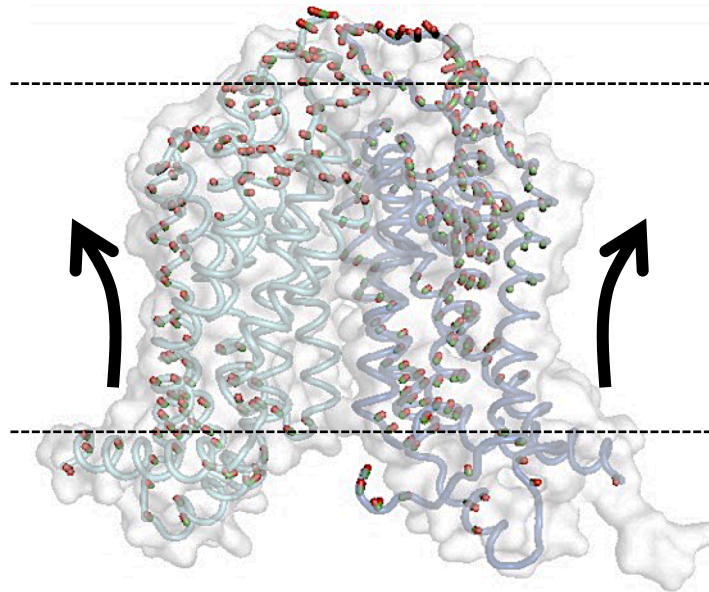
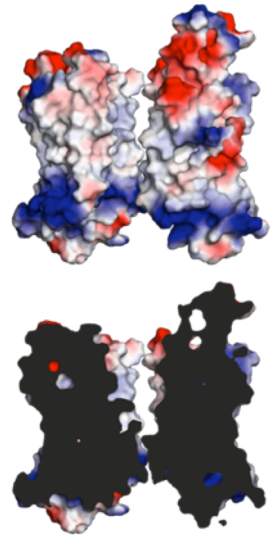
# Transmembrane Domain: the Dynamic Nature (Probably at an ATP-Prehydrolytic state)



**ABCG5/G8**  
(nf, inward)

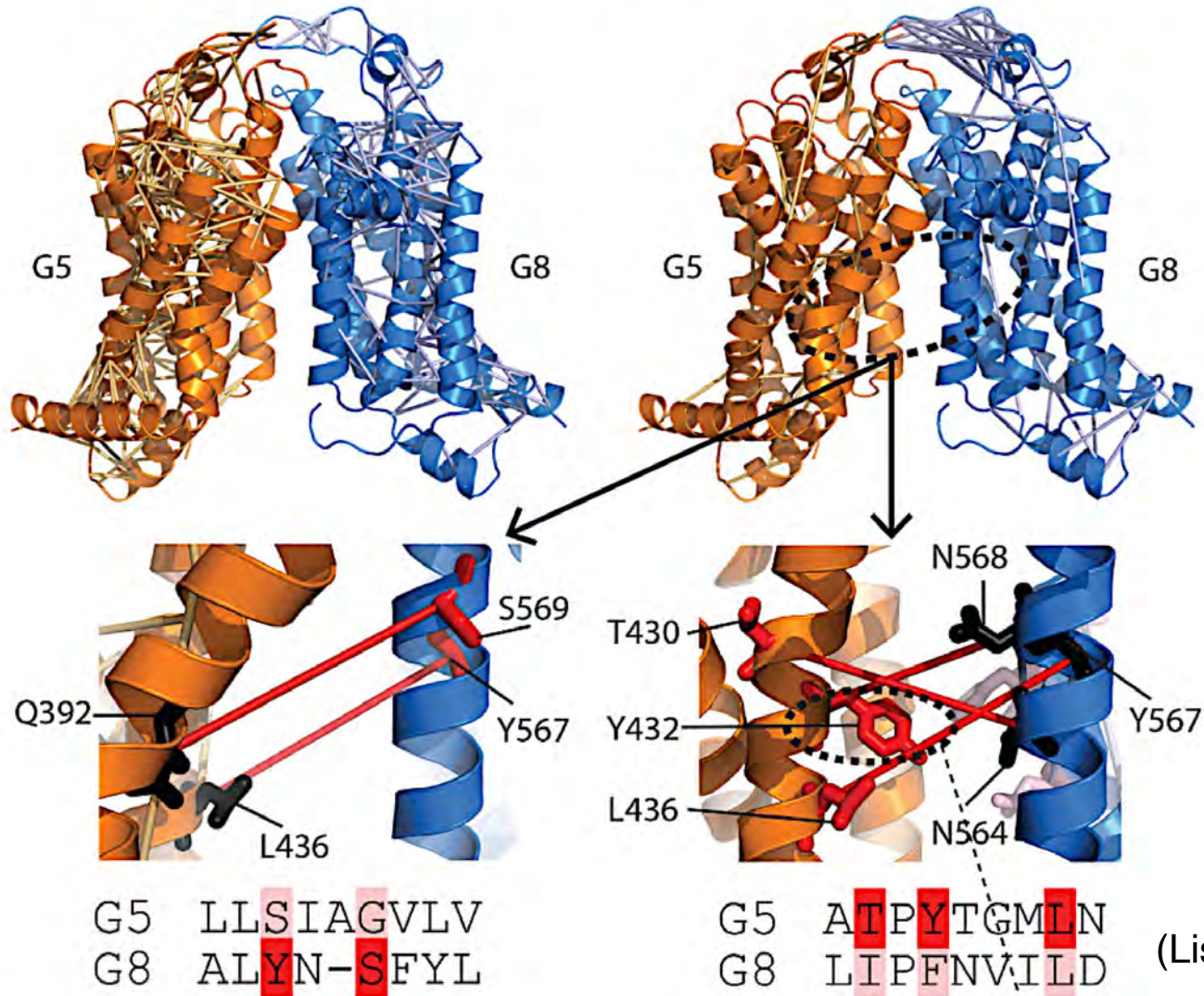


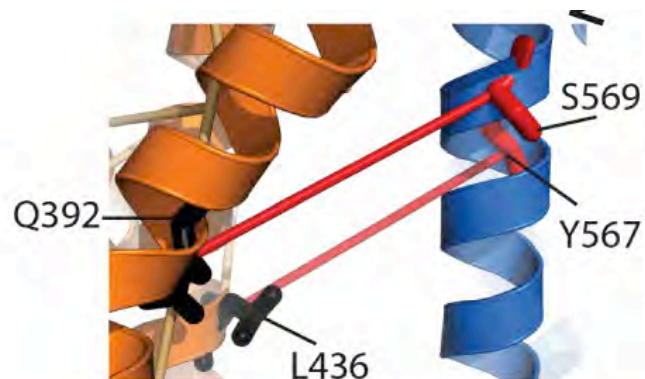
**ABCA1**  
(nf, outward)



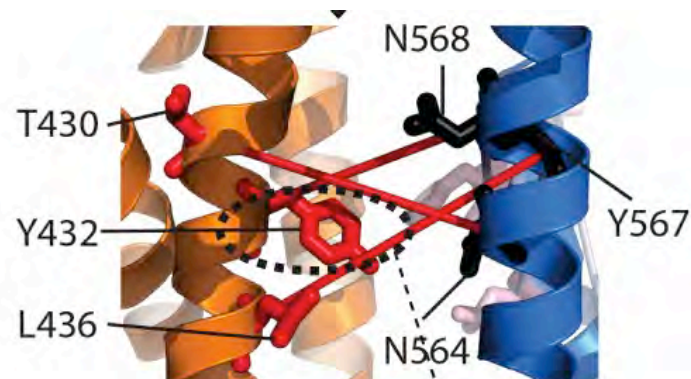
# Co-Evolution Analysis

- a** Coevolving residue pairs:  $\leq 8 \text{ \AA}$   
(within respective TMD)
- b** Coevolving residue pairs:  $> 8 \text{ \AA}$   
(candidate protein interface residues)



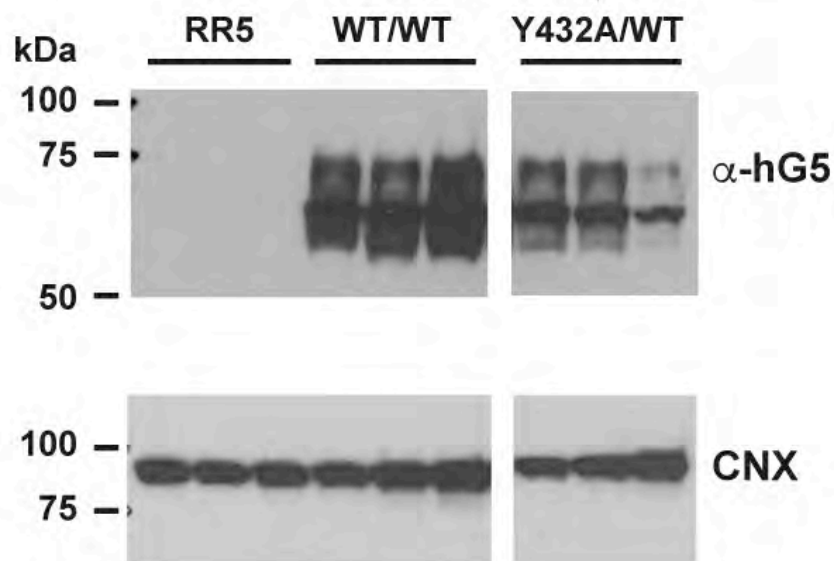
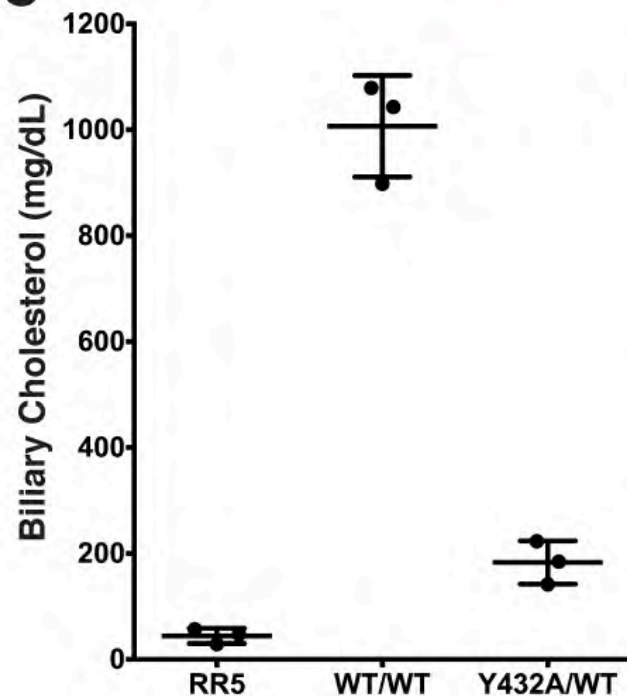


G5 LLSIAGVLV  
G8 ALYN-SFYL



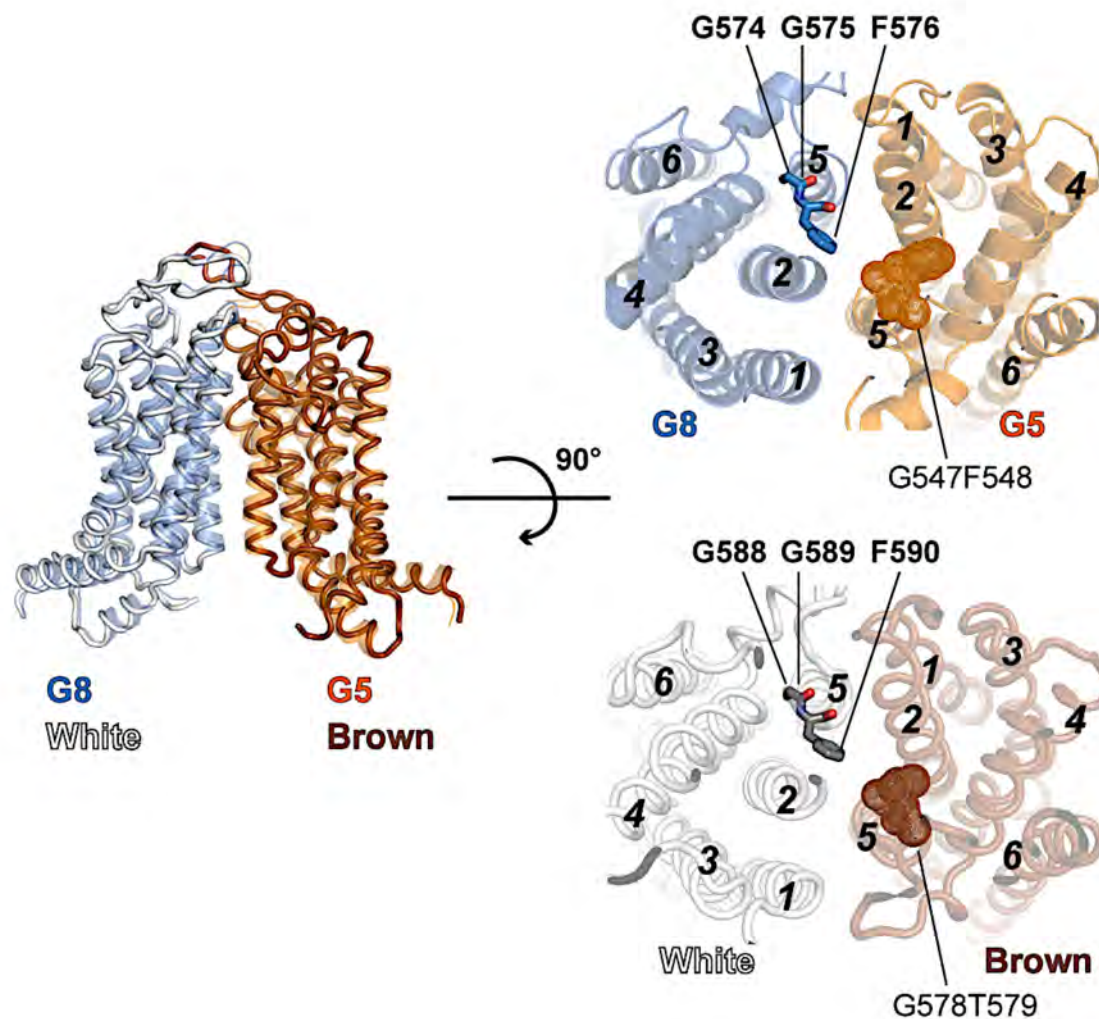
G5 ATPYTGMLN  
G8 LIPFNVILD

**C**



(Jin Wang & Fang Xu)



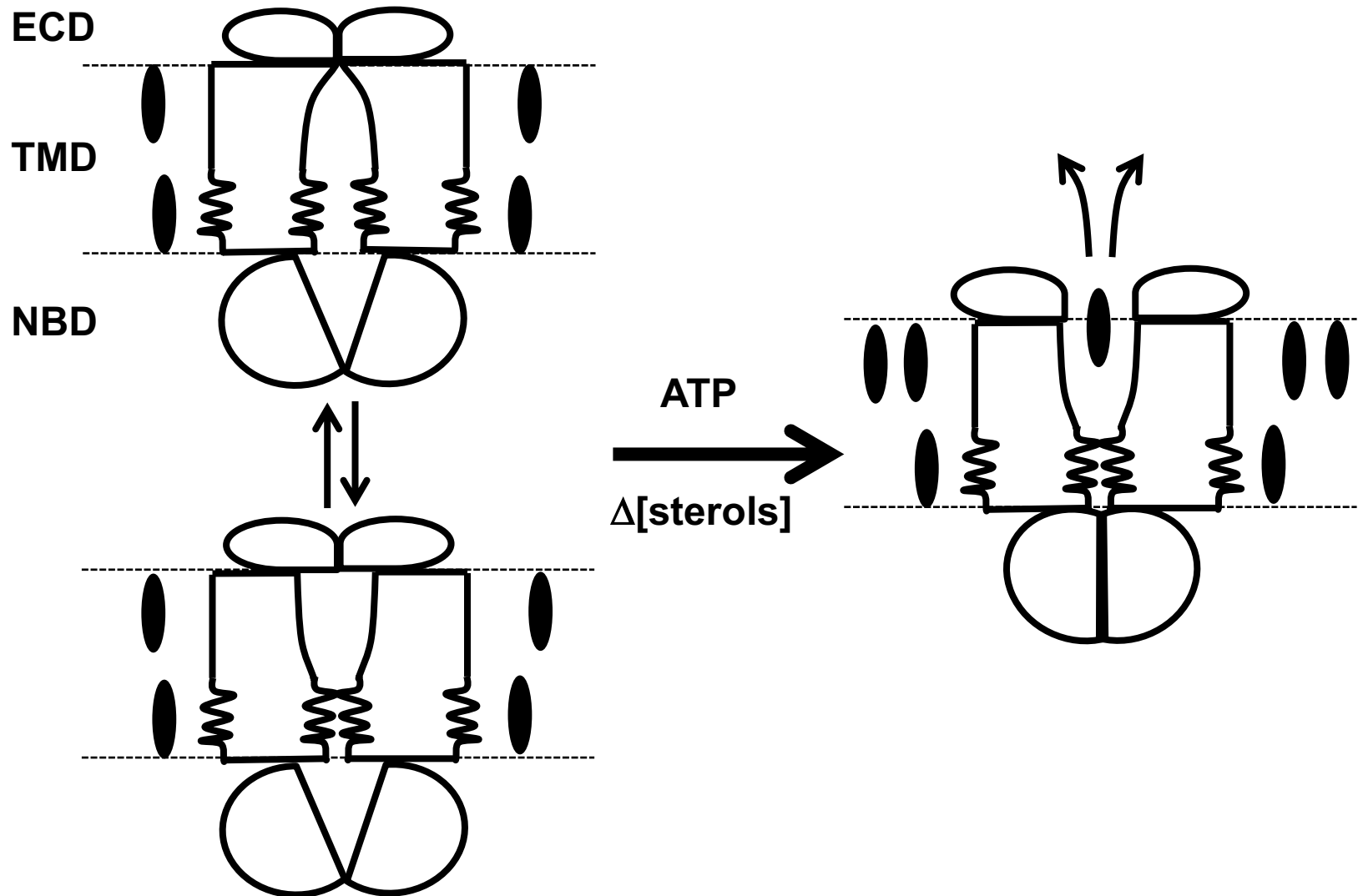


#### TMH5

<i>G5</i> ( <i>Homo sapiens</i> )	527	PNIVNSVVALLSIAGVLVGSGFLRN	551
<i>G5</i> ( <i>Danio rerio</i> )	531	PNMVNSGVALLNIAGIMVGSGFLRG	555
<i>G8</i> ( <i>Homo sapiens</i> )	556	FHMASFFSNALYN-SFYLAGGFMIN	579
<i>G8</i> ( <i>Danio rerio</i> )	537	LQTSSFMGNALFT-VFYLTAGFVIS	560
White	570	TSMALSVGPPVII-PFLLF <b>G</b> GFFLN	593
Brown	559	DKMASECAAPFDL-IFLIFGGTYMN	582
Scarlet	550	VPLAMAYLVPLDY-IFMITSGIFIQ	573

\*

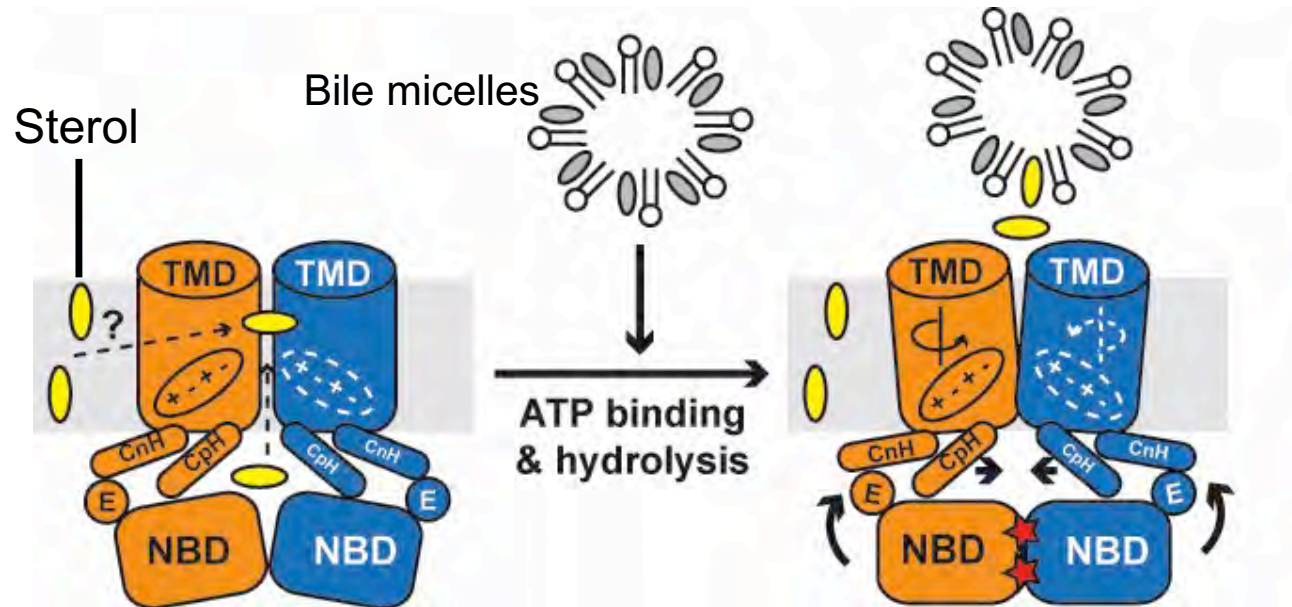
# Working Model of ABC Sterol Transporters (Cellular)



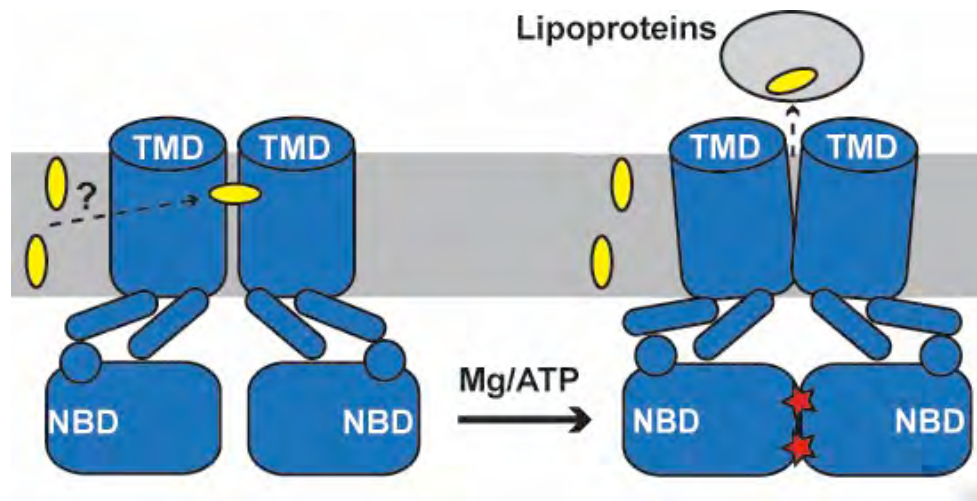


# Working Model of ABC Sterol Transporters (Molecular)

**A**



**B**



## So, ...

- **High-degree of structural diversity in the transmembrane domains of ABC transporters.**
- **The structural variability (likely) determines the functional diversity of ABC transporters.**
- **Transport mechanism is (likely) individually distinct.**